Title: NOVEL COMPOUNDS FOR MODULATING CELL PROLIFERATION

CROSS REFERENCE TO RELATED APPLICATIONS

This application claims the priority benefit of provisional application U.S. Serial No. 60/196,936, filed April 13, 2000, the contents of which are incorporated herein by reference in their entirety.

FIELD OF THE INVENTION

This invention relates to novel compounds which are useful for treating a variety of cell proliferative disorders such as cancer.

BACKGROUND OF THE INVENTION

A wide range of growth factors coordinate cell proliferation and differentiation. Mallgnant cells arise as a result of a stepwise progression of events that include the unregulated expression of growth factors or components of their signaling pathways. Tyrosine phosphorylation events initiated by receptor, cytoplasmic and nuclear kinases and regulated by phosphatases are central to these processes. Mutation, hyper-activation, translocation and overexpression of protein tyrosine kinases are all associated with tumorigenesis. In addition to increasing proliferative rates and immortalizing cells, overexpression of tyrosine kinases can lead to morphological transformation and cause anchorage independence, contributing to the promotion of migratory ability and possibly the induction of metastases.

Certain compounds with structures based upon mimicry of ATP or phosphotyrosine have been shown to be effective kinase inhibitors. Those based upon phosphotyrosine have been demonstrated to be the more specific tyrosine kinase inhibitors. Because of their ability to inhibit tyrosine phosphorylation, these compounds may alter cell responses to growth factors or other process driven by tyrosine kinase activity, including unregulated growth as the result of tyrosine kinase overexpression, mutation, or translocation. Inhibition of tyrosine kinases occupying a central role in

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proliferative signaling pathways, or in pathways regulating cell cytoskeletal structure, even temporary or incomplete inhibition, may be sufficient to switch a cancerous cell from a proliferative cycle into programmed cell death, or apoptosis. Death by apoptosis is most often observed upon effective treatment with tyrosine kinase inhibitors.

Selective inhibition of specific tyrosine kinases offers a method of targeting cancerous cell growth with a high degree of specificity and minimal toxicity to normally growing cells and tissues. Thus, specific inhibitors of tyrosine kinases have great potential as clinical anti-cancer treatments. A number of small molecules which act as tyrosine kinase inhibitors have been identified. For example, certain phenyl acrylonitrile compounds have been described as tyrosine kinase inhibitors, effective to inhibit cell proliferation (see for example, US 5,891,917, US 5,217,999, US 5,773,476, US 5,935,993, US 5,656,655, US 5,677,329 and US 5,789,427).

Inhibition of tyrosine kinases offers one mechanism by which cell proliferation can be inhibited. One of skill in the art will appreciate that other mechanisms of inhibition may also be involved.

There is a need in the art to identify compounds that inhibit cell proliferation.

SUMMARY OF THE INVENTION

A number of novel compounds have now been identified that inhibit abnormal cell proliferation, for example cancer cell growth. The compounds do not inhibit the growth of normal cells.

Accordingly, the present invention includes compounds of Formula I and salts, solvates and hydrates thereof:

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wherein

R¹ and R² are each indep indentity selected from the group consisting of H, OH, C₁₋₆alkyl, C₁₋₆alkoxy, NH₂, NH-C₁₋₆alkyl, N(C₁₋₆alkyl)(C₁₋₆alkyl), SH, S-C₁₋₆alkyl, O-Si(C₁₋₈alkyl)(C₁₋₆alkyl)(C₁₋₆alkyl), NO₂, CF₃, OCF₃ and halo; R³ is selected from the group consisting of H, OH, C₁₋₆alkyl, C₁₋₆alkoxy, NH₂, NH-C₁₋₆alkyl, N(C₁₋₆alkyl)(C₁₋₆alkyl), SH, S-C₁₋₆alkyl, O-Si(C₁₋₆alkyl)(C₁₋₆alkyl), NO₂, halo and CH₂-S-(CH₂)_n Ar; R⁴ is selected from the group consisting of C(X)R⁵, SO₃Ar, NH₂, NH-C₁₋₆alkyl, N(C₁₋₆alkyl)(C₁₋₆alkyl), P(O)(OH)₂, P(O)(OC₁₋₆alkyl)₂, and C(NH₂)=C(CN)₂; X is selected from O,S, NH and N-C₁₋₆alkyl;

10 R⁵ is selected from the group consisting of NH₂, OH, NH(CH₂)_pAr, NH(CH₂)_pOH, (CH₂)_pOC₁₋₆alkyl, C₁₋₆alkoxy, NHNH₂, NHC(O)NH₂, NHC(O)C₁₋₆alkoxy, N-morpholino and N-pyrrolidino;

Ar is an aromatic or heteroaromatic group, unsubstituted or substituted with 1-4 substituents, independently selected from the group consisting of OH,

15 C₁₋₈alkyl, C₁₋₆alkoxy, NH₂, NH-C₁₋₆alkyl, N(C₁₋₆alkyl)(C₁₋₆alkyl), SH, S-C₁₋₆alkyl, NO₂, CF₃, OCF₃ and halo; n is 0 to 4; and p is 1-4.

The present invention further includes compounds of Formula II and salts, solvates and hydrates thereof:

wherein

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R¹ and R² are each independently selected from the group consisting of H, OH, C₁₋₆alkyl, C₁₋₆alkoxy, NH₂, NH-C₁₋₆alkyl, N(C₁₋₆alkyl)(C₁₋₆alkyl), SH, S-C₁₋₆alkyl, O-Si(C₁₋₆alkyl)(C₁₋₆alkyl)(C₁₋₆alkyl), NO₂, CF₃, OCF₃ and halo; R³ is selected from the group consisting of H, OH, C₁₋₆alkyl, C₁₋₆alkoxy, NH₂, NH-C₁₋₆alkyl, N(C₁₋₆alkyl)(C₁₋₆alkyl), SH, S-C₁₋₆alkyl,

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O-Si(C₁₋₆alkyl)(C₁₋₆alkyl)(C₁₋₆alkyl), NO₂, halo and CH₂-S-(CH₂)_n Ar, Ar is an aromatic or heteroaromatic group, unsubstituted or substituted with 1-4 substituents, independently selected from the group consisting of OH, C_{1-6} alkyl, C_{1-6} alkoxy, NH_2 , $NH-C_{1-6}$ alkyl, $N(C_{1-6}$ alkyl)(C_{1-6} alkyl), SH_1

S-C₁₋₈alkyl, NO₂, CF₃, OCF₃ and halo;

R⁶ is selected from the group consisting of Ar, OH and OC₁₋₆alkyl;

X is selected from O and S;

n is 0-4; and

The present invention also provides compounds of Formula III and p is 1-4. 10 salts, solvates and hydrates thereof:

$$R^1$$
 R^2
 R^3

III

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 ${\sf R}^1$ and ${\sf R}^2$ are each independently selected from the group consisting of H, OH, C_{1-6} alkyl, C_{1-6} alkoxy, NH₂, NH- C_{1-6} alkyl, N(C_{1-6} alkyl), SH, 15 S-C₁₋₆alkyl, O-Si(C₁₋₆alkyl)(C₁₋₆alkyl), NO₂, CF₃, OCF₃ and halo; R^3 is selected from the group consisting of H, OH, C_{1-6} alkyl, C_{1-6} alkoxy, NH₂, NH-C₁₋₆alkyl, N(C₁₋₈alkyl)(C₁₋₆alkyl), SH, S-C₁₋₈alkyl,

O-Si(C₁₋₆alkyl)(C₁₋₆alkyl)(C₁₋₆alkyl), NO₂, halo and CH₂-S-(CH₂)_nAr, Ar is an aromatic or heteroaromatic group, unsubstituted or substituted with 1-20 4 substituents, independently selected from the group consisting of OH, C_{1-6} alkyl, C_{1-6} alkoxy, NH_2 , $NH-C_{1-6}$ alkyl, $N(C_{1-6}$ alkyl)(C_{1-6} alkyl), SH, S-C₁₋₆alkyl, NO₂, CF₃, OCF₃ and halo;

R⁷ is selected from the group consisting of OH, NH₂ and OC₁₋₆alkyl; X is selected from O and S; and n is 0-4.

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According to another aspect of the present invention, there is provided a pharmac utical composition comprising a compound of the invention and a pharmaceutically acceptable carrier or diluent.

In accordance with a further aspect of the present invention, there is provided a method for modulating cell proliferation, preferably inhibiting cell proliferation comprising administering an effective amount of a compound of the invention to a cell or animal in need thereof. The invention also includes a use of a compound of the invention to modulate cell proliferation, preferably inhibit cell proliferation. The invention further includes a use of a compound of the invention to prepare a medicament to modulate cell proliferation, preferably inhibit cell proliferation.

In a preferred embodiment the present invention provides a method of inhibiting the proliferation of a cancer cell comprising administering an effective amount of a compound of the invention to a cell or animal in need thereof. The cancer cell treated may be any type of cancer including a leukemia, a lymphoma, myeloma, metastatic carcinoma, sarcoma or any other malignant transformation or any other malignancy. The invention also includes a use of a compound of the invention to modulate cancer cell proliferation, preferably inhibit cancer cell proliferation. The invention further includes a use of a compound of the invention to prepare a medicament to modulate cancer cell proliferation, preferably inhibit cancer cell proliferation.

In another aspect, the invention provides a method of modulating tyrosine kinase activity in a cell by administering an effective amount of a compound of the invention. In a further aspect, the invention provides a method of inhibiting tyrosine kinase activity in a cell by administering an effective amount of a compound of the invention. The present invention also provides a use of a compound of the invention to modulate, preferably inhibit, tyrosine kinase activity. The present invention further provides a use of a compound of the invention to prepare a medicament to modulate tyrosine kinase activity, preferably inhibit tyrosine kinase activity. It is appreciated that the inhibition of cell growth by the compounds of the invention may be effected by other mechanisms.

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Other features and advantages of the present invention will become apparent from the following detail d description. It should be understood, however, that the detailed description and the specific examples while indicating preferred embodiments of the invention are given by way of illustration only, since various changes and modifications within the spirit and scope of the invention will become apparent to those skilled in the art from this detailed description.

BRIEF DESCRIPTION OF THE DRAWINGS

The invention will now be described in relation to the drawings in which:

Figure 1 is a bar graph showing the effect of CR4 upon normal bone marrow differentiation in culture.

Figure 2 is a bar graph showing the killing of Philadelphia positive acute lymphoblastic leukemia by low-dose CR4 in culture.

Figure 3 is a bar graph showing the killing of Philadelphia positive Z119 Acute lymphoblastic leukemia cells by low-dose CR4 in culture.

Figure 4 is a bar graph showing the killing of AML-3 acute myeloid leukemia cells by low-dose CR4 in culture.

Figure 5 is a bar graph showing the killing of Ly-MN lymphoma cells by low-dose CR4 in culture.

Figure 6 is a bar graph showing the killing of primary juvenile myelomonocytic leukemia cells by CR4 in culture.

Figure 7 is a bar graph showing the killing of OCI-LY2 lymphoma cells by low-dose CR4 in culture.

Figure 8 is a bar graph showing the killing of Philadelphia positive ALL cells by CR17 and CR21 in culture.

Figure 9 is a bar graph showing the killing of Philadelphia positive ALL cells by CR17 and CR21 in culture.

Figure 10 is a bar graph showing the killing of Philadelphia positive ALL cells by CR24 in culture.

Figure 11 is a bar graph showing the killing of Philadelphia positive ALL cells by CR19 in culture.

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Figure 12 is bar graph showing the effect of CR19 on normal bone marrow differentiation in culture.

Figure 13 is a bar graph showing the effect of CR24, CR17 and CR21 on normal bone marrow differentiation.

Figure 14 is a bar graph showing the effect of *in vitro* purging of normal bone marrow with CR4.

Figure 15 is a bar graph showing the effect of *in vitro* purging of Z119 acute lymphoblastic leukemia with CR4.

Figure 16 is a bar graph showing the effect of *In vitro* purging of OCI-10 Ly2 lymphoma cells with CR4.

Figure 17 is a bar graph showing the effect of *in vitro* purging of OCI-AML-3 acute meyloid leukemia cells with CR4.

Figure 18 is a bar graph showing the effect of *in vitro* purging of Ramos B cell Burkitt's lymphoma cells with CR4.

Figure 19 is a bar graph showing the killing of HuNS1 multiple myeloma cells by low-dose CR4 in culture.

Figures 20A and B are graphs showing cell staining after in vivo treatment of Philadelphia positive acute lymphoblastic leukemia in NOD-SCID mice.

Figure 21 is a bar graph showing that the effect of *in vitro* purging of normal bone marrow with CR11.

Figure 22 is a bar graph showing that the effect of *in vitro* purging of Philadelphia positive acute lymphoblastic leukemia with CR11.

Figure 23 is an autoradiograph which shows Philadelphia (Ph+) ALL lines Z119 and Z181 (5x10⁶ cells/point) immunoprecipitated with Bcr-Abl antibody.

Figure 24 is an autoradiograph which shows Philadelphia (Ph+) ALL line Z119 (5x10⁶ cells/point) immunoprecipitated with Jak2 antibody.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

Throughout this application, various publications, patents, and published patent applications are referred to by an identifying citation. The disclosures of the publications, patents, and published patent applications

referenced in this application are hereby incorporated by reference into the present disclosure in their entirety.

I. Definitions

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The term "C₁₋₆alkyl" as used herein means, unless otherwise stated, straight and/or branched chain alkyl radicals containing from one to six carbon atoms and includes methyl, ethyl, propyl, isopropyl, t-butyl and the like.

The term "C₁₋₈alkoxy" as used herein means, unless otherwise stated, straight and/or branched chain alkoxy radicals containing from one to six carbon atoms and includes methoxy, ethoxy, propyoxyl, isopropyloxy, t-butoxy and the like.

The term "C₁₋₄alkyl" as used herein means, unless otherwise stated, straight and/or branched chain alkyl radicals containing from one to four carbon atoms and includes methyl, ethyl, propyl, isopropyl, t-butyl and the like.

The term "C₁₋₄alkoxy" as used herein means, unless otherwise stated, straight and/or branched chain alkoxy radicals containing from one to four carbon atoms and includes methoxy, ethoxy, propyoxyl, isopropyloxy, t-butoxy and the like.

The term "Ar" as used herein, means an unsubstituted or substituted aryl and/or heteroaryl group which, in the case of heteroaryl, may contain up to two heteroatoms, wherein the constituents are independently selected from the group consisting of OH, C₁₋₆alkyl, C₁₋₆alkoxy, NH₂, NH-C₁₋₆alkyl, N(C₁₋₆alkyl)(C₁₋₆alkyl), SH, S-C₁₋₆alkyl, NO₂, CF₃, OCF₃ and halo, and includes unsubstituted or substituted phenyl, furyl, thienyl, indolyl, naphthyl, quinolyl and the like.

The term "halo" as used herein means halogen and includes chloro, flouro, bromo, iodo and the like.

The term "pharmaceutically acceptable salt" means an acid addition salt or a basic addition salt which is suitable for or compatible with the treatment of patients.

The term "compound of the invention" as used herein includes any compound of the Formula I, II or III as defined herein (including all salts,

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solvates or hydrates thereof) as well as a specific compound designated herein as CR1, CR2, CR3, CR4, CR5, CR8, CR9, CR11, CR12, CR13, CR14, CR15, CR16, CR17, CR18, CR19, CR20, CR21, CR24, CR27, CR28, and CR29 (including all salts, solvates or hydrates thereof).

The term "pharmaceutically acceptable acid addition salt" as used herein means any non-toxic organic or inorganic salt of any base compounds represented by Formulae I, II and/or III or any of their intermediates. Illustrative inorganic acids which form suitable salts include hydrochloric, hydrobromic, sulfuric and phosphoric acids, as well as metal salts such as sodium monohydrogen orthophosphate and potassium hydrogen sulfate. Illustrative organic acids that form suitable salts include mono-, di-, and tricarboxylic acids such as glycolic, lactic, pyruvic, malonic, succinic, glutaric, fumaric, malic, tartaric, citric, ascorblc, maleic, benzoic, phenylacetic, cinnamic and salicylic acids, as well as sulfonic acids such as p-toluene sulfonic and methanesulfonic acids. Either the mono or di-acid salts can be formed, and such salts may exist in either a hydrated, solvated or substantially anhydrous form. In general, the acid addition salts of compounds of Formulae I, II and/or III are more soluble in water and various hydrophilic organic solvents, and generally demonstrate higher melting points in comparison to their free base forms. The selection of the appropriate salt will be known to one skilled in the art. Other non-pharmaceutically acceptable salts, e.g. oxalates, may be used, for example, in the isolation of compounds of Formulae I, Il and/or III for laboratory use, or for subsequent conversion to a pharmaceutically acceptable acid addition salt.

The term "pharmaceutically acceptable basic addition salt" as used herein means any non-toxic organic or inorganic base addition salt of any acid compounds represented by Formulae I, II and/or III or any of their intermediates. Illustrative inorganic bases which form suitable salts include lithium, sodium, potassium, calcium, magnesium or barium hydroxide. Illustrative organic bases which form suitable salts include aliphatic, alicyclic or aromatic organic amines such as methylamine, trimethylamine and picoline

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or ammonia. The selection of the appropriate salt will be known to a person skilled in the art.

The term "solvate" as used herein means a compound of Formulae I, II and/or III, or a pharmaceutically acceptable salt of a compound of Formulae I, II and/or III, wherein molecules of a suitable solvent are incorporated in the crystal lattice. A suitable solvent is physiologically tolerable at the dosage administered. Examples of suitable solvents are ethanol, water and the like. When water is the solvent, the molecule is referred to as a "hydrate".

The term an "effective amount" or a "sufficient amount " of an agent as used herein is that amount sufficient to effect beneficial or desired results, including clinical results, and, as such, an "effective amount" depends upon the context in which it is being applied. For example, in the context of administering an agent that inhibits cancer cell proliferation, an effective amount of an agent is, for example, an amount sufficient to achieve such a reduction in cancer cell proliferation as compared to the response obtained without administration of the agent.

As used herein, and as well understood in the art, "treatment" is an approach for obtaining beneficial or desired results, including clinical results. Beneficial or desired clinical results can include, but are not limited to, alleviation or amelioration of one or more symptoms or conditions, diminishment of extent of disease, stabilized (i.e. not worsening) state of disease, preventing spread of disease, delay or slowing of disease progression, amelioration or palliation of the disease state, and remission (whether partial or total), whether detectable or undetectable. "Treatment" can also mean prolonging survival as compared to expected survival if not receiving treatment.

"Palliating" a disease or disorder means that the extent and/or undesirable clinical manifestations of a disorder or a disease state are lessened and/or time course of the progression is slowed or lengthened, as compared to not treating the disorder.

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The term "modulate" as used herein includes the inhibition or suppression of a function or activity (such as cell proliferation) as well as the enhancement of a function or activity.

To "Inhibit" or "suppress" or "reduce" a function or activity, such as cancer cell proliferation, is to reduce the function or activity when compared to otherwise same conditions except for a condition or parameter of interest, or alternatively, as compared to another conditions.

The term "animal" as used herein includes all members of the animal kingdom including human. The animal is preferably a human.

The term "a cell" as used herein includes a plurality of cells. Administering a compound to a cell includes in vivo, ex vivo and in vitro treatment.

The term "cancer cells" as used herein includes all forms of cancer or neoplastic disease.

15 II. Compounds of the Invention

Novel compounds which are useful in modulating cell proliferation were prepared. As such the compounds are useful in treating cell proliferative diseases such as cancer.

Accordingly, the present invention provides compounds of Formula I, and salts, solvates or hydrates thereof:

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wherein

R¹ and R² are each independently selected from the group consisting of H, OH, C₁₋₆alkyl, C₁₋₆alkoxy, NH₂, NH-C₁₋₆alkyl, N(C₁₋₆alkyl)(C₁₋₆alkyl), SH, S-C₁₋₆alkyl, O-Si(C₁₋₆alkyl)(C₁₋₆alkyl)(C₁₋₆alkyl), NO₂, CF₃, OCF₃ and halo; R³ is selected from the group consisting of H, OH, C₁₋₆alkyl, C₁₋₆alkoxy, NH₂, NH-C₁₋₆alkyl, N(C₁₋₆alkyl)(C₁₋₆alkyl), SH, S-C₁₋₆alkyl,

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O-Si(C₁₋₆alkyl)(C₁₋₆alkyl)(C₁₋₆alkyl), NO₂, halo and CH₂-S-(CH₂)_n Ar; R⁴ is selected from the group consisting of C(X)R⁵, SO₃Ar, NH₂, NH-C₁₋₆alkyl, N(C₁₋₆alkyl)(C₁₋₆alkyl), P(O)(OH)₂, P(O)(OC₁₋₆alkyl)₂, and C(NH₂)=C(CN)₂; X is selected from O,S, NH and N-C₁₋₆alkyl;

R⁵ is selected from the group consisting of NH₂, OH, NH(CH₂)_pAr, NH(CH₂)_pOH, (CH₂)_pOC₁₋₆alkyl, C₁₋₆alkyl, C₁₋₆alkoxy, NHNH₂, NHC(O)NH₂, NHC(O)C₁₋₆alkoxy, N-morpholino and N-pyrrolidino; and Ar is an aromatic or heteroaromatic group, unsubstituted or substituted with 1-4 substituents independently selected from the group consisting of OH,

C₁₋₆alkyl, C₁₋₆alkoxy, NH₂, NH-C₁₋₆alkyl, N(C₁₋₆alkyl)(C₁₋₆alkyl), SH, S-C₁₋₆alkyl, NO₂, CF₃, OCF₃ and halo; n is 0 to 4; m is 1 to 4; and p is 1-4.

In embodiments of the invention, compounds of Formula I are those in which R¹ and R² are each independently selected from the group consisting of H, OH, C₁-6alkyl, C₁-6alkoxy, NH₂, NH-C₁-6alkyl, N(C₁-6alkyl)(C₁-6alkyl), SH, S-C₁-6alkyl, O-Si(C₁-6alkyl)(C₁-6alkyl)(C₁-6alkyl), NO₂, CF₃, OCF₃ and halo. In preferred embodiments, R¹ and R² are each independently selected from the group consisting of H, OH, C₁-4alkyl, C₁-4alkoxy, NH₂, NH-C₁-4alkyl, SH, S-C₁-4alkyl, O-Si(C₁-4alkyl)(C₁-4alkyl), NO₂, CF₃, OCF₃ and halo. In more perferred embodiments, R¹ and R² are each independently selected from the group consisting of H, OH, OCH₃, O-Si(CH₃)₂(¹Bu), S-Me, SH and NO₂. In the most preferred embodiment of the present invention R¹ and R² are both OH or OCH₃ or R¹ is OCH₃ and R² is OH.

In further embodiments of the present invention, the compounds of Formula I include those in which R³ is selected from the group consisting of H, OH, C₁₋₆alkyl, C₁₋₆alkoxy, NH₂, NH-C₁₋₆alkyl, N(C₁₋₆alkyl)(C₁₋₆alkyl), SH, S-C₁₋₆alkyl, O-Si(C₁₋₆alkyl)(C₁₋₆alkyl)(C₁₋₆alkyl), NO₂, halo and CH₂-S-(CH₂)_n Ar (where n is 0-4). In preferred embodiments, R³ is selected from the group consisting of H, OH, C₁₋₄alkyl, C₁₋₄alkoxy, NH₂, NH-C₁₋₄alkyl, N(C₁₋₄alkyl)(C₁₋₄alkyl), SH, S-C₁₋₄alkyl, NO₂ and halo. In a more preferred embodiment, R³ is

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selected from the group consisting of H, OH, OCH₃, SH, SMe, NO₂ and halo. In the most preferred embodment, R^3 is selected from the group consisting of H, OH and OCH₃.

Embodiments of the invention include compounds of Formula I wherein R^4 is selected from the group consisting of $C(X)R^5$, SO_3Ar , NH_2 , $NH-C_{1-6}alkyl$, $N(C_{1-6}alkyl)(C_{1-6}alkyl)$, $P(O)(OH)_2$, $P(O)(OC_{1-6}alkyl)_2$, and $C(NH_2)=C(CN)_2$ (where m is 1-4). In preferred embodiments, R^4 is selected from the group consisting of $C(X)R^5$ and $C(NH_2)=C(CN)_2$. More preferably, R^4 is $C(X)R^5$. When R^4 is $C(X)R^5$, embodiments of the invention include compounds where X is selected from O,S, NH and N-C₁₋₆alkyl and R^5 is selected from the group consisting of NH_2 , OH, $NH(CH_2)_pAr$, $NH(CH_2)_pOH$, $(CH_2)_pOC_{1-6}alkyl$, $C_{1-6}alkyl$, $C_{1-6}alkoxy$, $NHNH_2$, $NHC(O)NH_2$, $NHC(O)C_{1-6}alkoxy$, N-morpholino and N-pyrrolidino (where p is 1-4). In preferred embodiments, X is O or S and R^5 is selected from the group consisting of NH_2 , OH, $NH(CH_2)_pAr$, $(CH_2)_pOH$ and $C_{1-4}alkoxy$, (where p is 1-3). Most preferred, are compounds of Formula I wherein X is O and R^5 is selected from the group consisting of NH_2 , OH, $NH(CH_2)_pAr$, $NH(CH_2)_pOH$ and OCH_3 , (where p is 1-2).

The present invention includes compounds of Formula I wherein the term "Ar" means an unsubstituted or substituted aryl and/or heteroaryl group which, in the case of heteroaryl, may contain up to two heteroatoms, wherein the optional substituents are independently selected from the group consisting of OH, C₁₋₈alkyl, C₁₋₈alkoxy, NH₂, NH-C₁₋₆alkyl, N(C₁₋₈alkyl)(C₁₋₆alkyl), SH, S-C₁₋₆alkyl, NO₂, CF₃, OCF₃ and halo, and includes unsubstituted or substituted phenyl, furyl, thienyl, indolyl, naphthyl, quinolyl and the like. In embodiments of the present invention, Ar is an unsubstituted phenyl group or a phenyl group substituted with 1-4 substituents optionally selected from the group consisting of OH, C₁₋₆alkyl, C₁₋₆alkoxy, NH₂, NH-C₁₋₆alkyl, N(C₁₋₆alkyl)(C₁₋₆alkyl), SH, S-C₁₋₆alkyl, NO₂, CF₃, OCF₃ and halo. In preferred embodiments, Ar is an unsubstituted phenyl group or phenyl group substituted with 1-2 substituents optionally selected from the group consisting of OH, C₁₋₄alkyl, C₁₋₄alkoxy, NH₂, NH-C₁₋₄alkyl, N(C₁₋₄alkyl), SH, S-C₁₋₄alkyl, C₁₋₄alkoxy, NH₂, NH-C₁₋₄alkyl, N(C₁₋₄alkyl), SH, S-C₁₋₄alkyl, NO₂, CF₃, OCF₃ and halo. In more preferred embodiments, Ar is an

unsubstituted phenyl group or phenyl group substituted with 1-2 substituents optionally select d from the group consisting of OH, OCH₃, NH₂, NHCH₃, N(CH₃)₂, SH, SCH₃, CF₃, OCF₃ and halo. In the most preferred embodiment, Ar is selected from the group consisting of phenyl and 3,4-dihydroxyphenyl.

The present invention further includes compounds of Formula II and salts, solvates and hydrates thereof:

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R¹ and R² are each independently selected from the group consisting of H, OH, C_{1-6} alkyl, C_{1-6} alkoxy, NH_2 , $NH-C_{1-6}$ alkyl, $N(C_{1-6}$ alkyl)(C_{1-6} alkyl), SH, S- C_{1-6} alkyl, O-Si(C_{1-6} alkyl)(C_{1-6} alkyl)(C_{1-6} alkyl), NO_2 , CF_3 , OCF_3 and halo; R³ is selected from the group consisting of H, OH, C_{1-6} alkyl, C_{1-6} alkoxy, NH_2 ,

NH-C₁₋₆alkyl, N(C₁₋₆alkyl)(C₁₋₆alkyl), SH, S-C₁₋₆alkyl,
O-Si(C₁₋₆alkyl)(C₁₋₆alkyl)(C₁₋₆alkyl), NO₂, halo and CH₂-S-(CH₂)_n Ar;
Ar is an aromatic or heteroaromatic group, unsubstituted or substituted with 1-4 substituents, independently selected from the group consisting of OH,
C₁₋₆alkyl, C₁₋₆alkoxy, NH₂, NH-C₁₋₆alkyl, N(C₁₋₆alkyl)(C₁₋₆alkyl), SH,

S-C₁₋₆alkyl, NO₂, CF₃, OCF₃ and halo;

R⁶ is selected from the group consisting of Ar, OH and OC₁₋₆alkyl;

X is selected from O and S;

n is 0-4; and

p is 1-4.

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In embodiments of the invention, compounds of Formula II are those in which R¹ and R² are each independently selected from the group consisting of H, OH, C₁₋₆alkyl, C₁₋₆alkoxy, NH₂, NH-C₁₋₆alkyl, N(C₁₋₆alkyl)(C₁₋₆alkyl), SH, S-C₁₋₆alkyl, O-Si(C₁₋₆alkyl)(C₁₋₆alkyl), NO₂, CF₃, OCF₃ and halo. In

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preferred embodiments, R^1 and R^2 are each Independently selected from the group consisting of H, OH, C_{1-4} alkyl, C_{1-4} alkoxy, NH_2 , $NH-C_{1-4}$ alkyl, SH, S- C_{1-4} alkyl, O-Si(C_{1-4} alkyl)(C_{1-4} alkyl), NO_2 , CF_3 , OCF_3 and halo. In more perferred embodiments, R^1 and R^2 are each independently selected from the group consisting of H, OH, OCH_3 , $O-Si(CH_3)_2(^tBu)$, S-Me, SH and NO_2 . In the most preferred embodiment of the present invention R^1 and R^2 are both OH or OCH_3 or R^1 is OCH_3 and R^2 is OH.

In further embodiments of the present invention, the compounds of Formula II include those in which R³ is selected from the group consisting of H, OH, C₁₋₆alkyl, C₁₋₆alkoxy, NH₂, NH-C₁₋₆alkyl, N(C₁₋₆alkyl)(C₁₋₆alkyl), SH, S-C₁₋₆alkyl, O-Si(C₁₋₆alkyl)(C₁₋₆alkyl)(C₁₋₆alkyl), NO₂, halo and CH₂-S-(CH₂)_n Ar (where n is 0-4). In preferred embodiments, R³ is selected from the group consisting of H, OH, C₁₋₄alkyl, C₁₋₄alkoxy, NH₂, NH-C₁₋₄alkyl, N(C₁₋₄alkyl)(C₁₋₄alkyl), SH, S-C₁₋₄alkyl, NO₂ and halo. In a more preferred embodiment, R³ is selected from the group consisting of H, OH, OCH₃, SH, SMe, NO₂ and halo. In the most preferred embodiment, R³ is selected from the group consisting of H, OH and OCH₃.

The present invention further includes compounds of Formula II wherein the term "Ar" means an unsubstituted or substituted aryl and heteroaryl group which, in the case of heteroaryl, may contain up to two heteroatoms, wherein the optional substituents are independently selected from the group consisting of OH, C₁₋₆alkyl, C₁₋₆alkoxy, NH₂, NH-C₁₋₆alkyl, N(C₁₋₆alkyl)(C₁₋₆alkyl), SH, S-C₁₋₆alkyl, NO₂, CF₃, OCF₃ and halo, and includes unsubstituted or substituted phenyl, furyl, thienyl, indolyl, naphthyl, quinolyl and the like. In embodiments of the present invention, Ar is an unsubstituted phenyl group or a phenyl group substituted with 1-4 substituents optionally selected from the group consisting of OH, C₁₋₆alkyl, C₁₋₆alkoxy, NH₂, NH-C₁₋₆alkyl, N(C₁₋₆alkyl)(C₁₋₆alkyl), SH, S-C₁₋₆alkyl, NO₂, CF₃, OCF₃ and halo. In preferred embodiments, Ar is an unsubstituted phenyl group or phenyl group substituted with 1-2 substituents optionally selected from the group consisting of OH, C₁₋₄alkyl, C₁₋₄alkoxy, NH₂, NH-C₁₋₄alkyl, N(C₁₋₄alkyl), SH, S-C₁₋₄alkyl, NO₂, CF₃, OCF₃ and halo. In more

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pr ferred embodiments, Ar is an unsubstituted phenyl group or ph nyl group substituted with 1-2 substituents optionally selected from the group consisting of OH, OCH₃, NH₂, NHCH₃, N(CH₃)₂, SH, SCH₃, CF₃, OCF₃ and halo. In the most preferred embodiment, Ar is selected from the group consisting of phenyl and 3,4-dihydroxyphenyl.

The compounds of Formula II, include those in which R⁶ is selected from the group consisting of Ar, OH and OC1-6alkyl and p is 1-4. In preferred embodiments, R⁶ is selected from the group consisting of Ar and OH and p is 1-2. Most preferably, when R⁶ is Ar, p is 1 and when R⁶ is OH, p is 2. Where R⁶ is Ar, Ar means an unsubstituted or substituted aryl and/or heteroaryl group which, in the case of heteroaryl, may contain up to two heteroatoms, wherein the optional substituents are independently selected from the group consisting of OH, C₁₋₆alkyl, C₁₋₆alkoxy, NH₂, NH-C₁₋₆alkyl, N(C₁₋₆alkyl)(C₁. 6alkyl), SH, S-C₁₋₆alkyl, NO₂, CF₃, OCF₃ and halo, and includes unsubstituted or substituted phenyl, furyl, thienyl, indolyl, naphthyl, quinolyl and the like. In embodiments of the present invention, Ar is an unsubstituted phenyl group or a phenyl group substituted with 1-4 substituents optionally selected from the group consisting of OH, C₁₋₆alkyl, C₁₋₆alkoxy, NH₂, NH-C₁₋₆alkyl, N(C₁₋₆alkyl, N(C₁₋₆ 6alkyl)(C1-6alkyl), SH, S-C1-6alkyl, NO2, CF3, OCF3 and halo. In preferred embodiments, Ar is an unsubstituted phenyl group or phenyl group substituted with 1-2 substituents optionally selected from the group consisting of OH, C1. 4alkyl, C1-4alkoxy, NH2, NH-C1-4alkyl, N(C1-4alkyl)(C1-4alkyl), SH, S-C1-4alkyl, NO₂, CF₃, OCF₃ and halo. In more preferred embodiments, Ar is an unsubstituted phenyl group or phenyl group substituted with 1-2 substituents optionally selected from the group consisting of OH, OCH₃, NH₂, NHCH₃, N(CH₃)₂, SH, SCH₃, CF₃, OCF₃ and halo. In the most preferred embodiment, Ar is selected from the group consisting of phenyl and 3,4-dihydroxyphenyl.

Compounds of Formula II, further include those in which X is selected from O and S. In preferred embodiments, X is O.

The present invention also provides a compound of Formula III and salts, solvates and hydrates thereof:

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wherein

R¹ and R² are each independently selected from the group consisting of H, OH, C₁₋₆alkyl, C₁₋₆alkoxy, NH₂, NH-C₁₋₆alkyl, N(C₁₋₆alkyl)(C₁₋₆alkyl), SH,

S-C₁₋₆alkyl, O-Si(C₁₋₆alkyl)(C₁₋₆alkyl)(C₁₋₆alkyl), NO₂, CF₃, OCF₃ and halo; R³ is selected from the group consisting of H, OH, C₁₋₆alkyl, C₁₋₆alkoxy, NH₂, NH-C₁₋₆alkyl, N(C₁₋₆alkyl)(C₁₋₆alkyl), SH, S-C₁₋₆alkyl,

O-Si(C₁₋₆alkyl)(C₁₋₆alkyl)(C₁₋₆alkyl), NO₂, halo and CH₂-S-(CH₂)_n Ar;

Ar is an aromatic or heteroaromatic group, unsubstituted or substituted with 1-

4 substituents, independently selected from the group consisting of OH,

 C_{1-6} alkyl, C_{1-6} alkoxy, NH_2 , $NH-C_{1-6}$ alkyl, $N(C_{1-6}$ alkyl), C_{1-6} alkyl), SH,

S-C₁₋₆alkyl, NO₂, CF₃, OCF₃ and halo;

R⁷ is selected from the group consisting of OH, NH₂ and OC₁₋₆alkyl;

X is selected from O and S; and

15 n is 0-4.

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In embodiments of the invention, compounds of Formula III are those in which R¹ and R² are each independently selected from the group consisting of H, OH, C₁-6alkyl, C₁-6alkoy, NH₂, NH-C₁-6alkyl, N(C₁-6alkyl)(C₁-6alkyl), SH, S-C₁-6alkyl, O-Si(C₁-6alkyl)(C₁-6alkyl)(C₁-6alkyl), NO₂, CF₃, OCF₃ and halo. In preferred embodiments, R¹ and R² are each independently selected from the group consisting of H, OH, C₁-4alkyl, C₁-4alkoxy, NH₂, NH-C₁-4alkyl, SH, S-C₁-4alkyl, O-Si(C₁-4alkyl)(C₁-4alkyl), NO₂, CF₃, OCF₃ and halo. In more perferred embodiments, R¹ and R² are each independently selected from the group consisting of H, OH, OCH₃, O-Si(CH₃)₂(¹Bu), S-Me, SH and NO₂. In the most preferred embodiment of the present invention R¹ and R² are both OH or OCH₃ or R¹ is OCH₃ and R² is OH.

In further embodiments of the present invention, the compounds of Formula III include those in which R³ is selected from the group consisting of

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H, OH, C₁₋₆alkyl, C₁₋₆alkoxy, NH₂, NH-C₁₋₆alkyl, N(C₁₋₆alkyl)(C₁₋₆alkyl), SH, S-C₁₋₆alkyl, O-SI(C₁₋₆alkyl)(C₁₋₆alkyl)(C₁₋₆alkyl), NO₂, halo and CH₂-S-(CH₂)_n Ar (where n is 0-4). In preferred embodiments, R³ is selected from the group consisting of H, OH, C₁₋₄alkyl, C₁₋₄alkoxy, NH₂, NH-C₁₋₄alkyl, N(C₁₋₄alkyl)(C₁₋₄alkyl), SH, S-C₁₋₄alkyl, NO₂ and halo. In a more preferred embodiment, R³ is selected from the group consisting of H, OH, OCH₃, SH, SMe, NO₂ and halo. In the most preferred embodment, R³ is selected from the group consisting of H, OH and OCH₃.

The present invention further includes compounds of Formula III wherein the term "Ar" means an unsubstituted or substituted aryl and/or heteroaryl group which, in the case of heteroaryl, may contain up to two heteroatoms, wherein the optional substituents are independently selected from the group consisting of OH, C₁₋₆alkyl, C₁₋₆alkoxy, NH₂, NH-C₁₋₆alkyl, N(C₁₋₆alkyl)(C₁₋₆alkyl), SH, S-C₁₋₆alkyl, NO₂, CF₃, OCF₃ and halo, and includes unsubstituted or substituted phenyl, furyl, thienyl, indolyl, naphthyl, quinolyl and the like. In embodiments of the present invention, Ar is an unsubstituted phenyl group or a phenyl group substituted with 1-4 substituents optionally selected from the group consisting of OH, C₁₋₆alkyl, C₁₋ $_{6}$ alkoxy, NH₂, NH-C₁₋₆alkyl, N(C₁₋₆alkyl)(C₁₋₆alkyl), SH, S-C₁₋₆alkyl, NO₂, CF₃, OCF₃ and halo. In preferred embodiments, Ar is an unsubstituted phenyl group or phenyl group substituted with 1-2 substituents optionally selected from the group consisting of OH, C₁₋₄alkyl, C₁₋₄alkoxy, NH₂, NH-C₁₋₄alkyl, N(C₁₋₄alkyl)(C₁₋₄alkyl), SH, S-C₁₋₄alkyl, NO₂, CF₃, OCF₃ and halo. In more preferred embodiments, Ar is an unsubstituted phenyl group or phenyl group substituted with 1-2 substituents optionally selected from the group consisting of OH, OCH₃, NH₂, NHCH₃, N(CH₃)₂, SH, SCH₃, CF₃, OCF₃ and halo. In the most preferred embodiment, Ar is selected from the group consisting of phenyl and 3,4-dihydroxyphenyl.

Compounds of Formula III further include those in which R^7 is selected from the group consisting of OH, NH_2 and OC_{1-6} alkyl. In preferred embodiments, R^7 is selected from the group consisting of OH and NH_2 .

Compounds of Formula III, further include those in which X is selected from O and S. In preferred embodiments, X is O.

In specific embodiments of the present invention, the compounds of the invention include:

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5 (E,E)-2-(benzylamido)-3-styrylacrylonitrile (CR1);
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- (E,E)-2-(benzylamido)-3-(3,4-dimethoxystyryl)acrylonitrile (CR2);
- (E,E)-2-(benzylamido)-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR3);
- (E,E)-2-(benzylamido)-3-(3,4-dihydroxystyryl)acrylonitrile (CR4);
- (E,E)-2-(phenylethylamido)-3-(3,4-dimethoxystyryl)acrylonitrile (CR5);
- (E,E)-2-(phenylethylamido)-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR8);
 - (E,E)-2-(phenylpropylamido)-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR9);
 - (E,E)-2-(3,4-dihydroxybenzylamido)-3-(3,5-dimethoxy-4-
- 15 hydroxystyryl)acrylonitrile (CR11);
 - (E,E)-2-thioacetamido-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR12);
 - (E,E)-2-acetamido-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR13);
 - (E,E)-2-carboxy-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR14);
 - (E,E)-2-carbomethoxy-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR15);
- 20 (E,E)-2-acetamido-3-[3,4-bis(t-butyldimethylsilyloxystyryl)]acrylonitrile(CR16);
 - (E,E)-2-acetamido-3-(3,4-dihydroxystyryl)acrylonitrile (CR17);
 - (E,E)-2-(benzylamido)-3-(3,4-bis(t-butyldimethylsilyloxystyryl))acrylonitrile (CR18);
 - (E,E)-2-(3,4 dihydroxybenzylamido)-3-styrylacrylonitrile (CR19);
- 25 (E,E)-2-(3,4 dihydroxybenzylamido)-3-[3,4-bis(t
 - butyldimethylsilyloxystyryl)]acrylonitrile (CR20);
 - (E,E)-2-(3,4 dihydroxybenzylamido)-3-(3,4-dihydroxystyryl)acrylonitrile (CR21);
- (E,E)-2-(β -ethanolamido)-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile 30 (CR24);
 - (E,E)-2-(benzylamido)-3-(4-nitrostyryl)acrylonitrile (CR27);
 - (E,E)-2-(3,4-dihydroxybenzylamido)-3-(4-nitrostyryl)acrylonitrile(CR28); and

(E,E)-2-(1-amino-2,2-dicyanoethenyl)-3-(4-nitrostyryl)acrylonitrile (CR29).

In preferred embodiments of the present invention, the compounds of the invention include:

- (E,E)-2-(benzylamido)-3-styrylacrylonitrile (CR1);
- 5 (E,E)-2-(benzylamido)-3-(3,4-dimethoxystyryl)acrylonitrile (CR2);
 - (E,E)-2-(benzylamido)-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR3);
 - (E,E)-2-(benzylamido)-3-(3,4-dihydroxystyryl)acrylonitrile (CR4);
 - (E,E)-2-(phenylethylamido)-3-(3,4-dimethoxystyryl)acrylonitrile (CR5);
 - (E,E)-2-(phenylpropylamido)-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile
- 10 (CR9);
 - (E,E)-2-(3,4-dihydroxybenzylamido)-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR11);
 - (E,E)-2-thioacetamido-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR12);
 - (E,E)-2-acetamido-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR13);
- 15 (E,E)-2-carboxy-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR14);
 - (E,E)-2-carbomethoxy-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR15);
 - (E,E)-2-acetamido-3-(3,4-dihydroxystyryl)acrylonitrile (CR17);
 - (E,E)-2-(3,4 dihydroxybenzylamido)-3-styrylacrylonitrile (CR19);
 - (E,E)-2-(3,4 dihydroxybenzylamido)-3-(3,4-dihydroxystyryl)acrylonitrile
- 20 (CR21); and
 - (E,E)-2- $(\beta$ -ethanolamido)-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR24).

In more preferred embodiments of the present invention, the compounds of the invention include:

- 25 (E,E)-2-(benzylamido)-3-(3,4-dihydroxystyryl)acrylonitrile (CR4);
 - (E,E)-2-(3,4-dihydroxybenzylamido)-3-(3,5-dimethoxy-4-

hydroxystyryl)acrylonitrile (CR11);

- (E,E)-2-acetamido-3-(3,4-dihydroxystyryl)acrylonitrile (CR17);
- (E,E)-2-(3,4 dihydroxybenzylamido)-3-styrylacrylonitrile (CR19);
- 30 (E,E)-2-(3,4 dihydroxybenzylamido)-3-(3,4-dihydroxystyryl)acrylonitrile (CR21); and

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(E,E)-2- $(\beta$ -ethanolamido)-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR24).

The present invention includes within its scope, prodrugs of the compounds of the invention. In general, such prodrugs will be functional derivatives of a compound of the invention which are readily convertible *in vivo* into the compound from which it is notionally derived. Conventional procedures for the selection and preparation of suitable prodrugs are described, for example, in "Design of Prodrugs" ed. H. Bundgaard, Elsevier, 1985.

Some of the compounds of the invention may have at least one asymmetric center. Where the compounds according to the invention have one asymmetric center, the may exist as enantiomers. Where the compounds of the invention possess two or more asymmetric centers, they may additionally exist as diastereomers. It is to be understood that all such isomers and mixtures thereof in any proportion are encompassed within the scope of the present invention.

The present invention includes radiolabeled forms of compounds of the invention, for example, compounds of the invention labeled by incorporation within the structure ³H or ¹⁴C or a radioactive halogen such as ¹²⁵I.

The compounds of the invention may, for example, be derived from an activated cinnamyl compound and an activated cyano-substituted methylene compound. A person skilled in the art, therefore, may wish to provide a generic name for the compounds of the invention based on the cinnamyl moiety. However, generic nomenclature based on the formed acylonitrile moiety, for example, styryl acrylonitrile, would be more proper.

III. Methods of Preparing Compounds of the Invention

In accordance with another aspect of the present invention, the compounds of the invention can be prepared by processes analogous to those established in the art. Therefore, compounds of this invention may be prepared by the reaction sequence shown in Scheme 1:

Scheme 1

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R1 CHO
$$\frac{R^4}{CN}$$
 $\frac{R^4}{(V)}$ $\frac{R^4}{EtOH, \beta-alanine}$ $\frac{R^4}{R^2}$ $\frac{R^4}{(I)}$ $\frac{R^4}{CN}$

Compounds of the general Formulae I, II and/or III useful in the practice of this invention can be prepared by Knoevenagel condensation of α,β -unsaturated aldehydes, such as cinnamaldehyde or its various arylsubstituted homologues (IV), with a compound having active α -methylene group (V). Similar Knoevenagel condensations using ylidenemalononitriles as active α -methylene group components were described in a review (F. Freeman. Chem. Rev. 1980, V. 80, P. 329-350). For example, these condensations may be carried out in a polar solvent, such as ethanol, in the presence of catalytic amounts of a weak base, such as β -alanine. Reaction temperatures may be in the range of 20 to 100°C, depending on the stability of the materials used in the condensation.

Compounds of Formulae IV and/or V may be commercially available, such as cinnamaldehyde, and its 3,5-dimethoxy-4-hydroxy derivative. Other compounds of Formulae IV and/or V may be prepared using straightforward procedures. For example, various R¹, R², R³-hydroxy substituted cinnamaldehydes can be prepared from the corresponding commercially available aryl substituted cinnamic acids. Scheme 2 gives an example of the preparation of protected 3,4-dihydroxycinnamaldehyde (IVa) starting from 3,4-dihydroxycinnamic acid (VI). At the end of the reaction sequence, the protection groups can be removed using standard methods well known to those having skill in the art.

Scheme 2

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R¹, R², R³ substituents may be also converted from one functional group to another, for example by known reduction of nitro groups into amino groups and the further transformation into dialkylamino groups, or by known conversion of hydroxy groups to halo groups.

α-Cyano amides with a reactive methylene group (Va) may be obtained, for example, as described in A. Gazit et.al. J. Med. Chem., 1991, V. 34, P. 1896-1907. For example, by heating methyl cyanoacetate (VII) and an appropriate commercially available amine (VIII) up to 100°C without presence of a solvent for 12-15 h followed by vacuum distillation directly from the mixture (for example using a Kugelrohr apparatus), the desired products may be obtained (Scheme 3).

Scheme 3

In some cases the chemistries outlined above may have to be modified, for instance by use of protective groups, to prevent side reactions due to reactive groups, such as reactive groups attached as substituents. This may be achieved by means of conventional protecting groups, for example as described in "Protective Groups in Organic Chemistry" McOmie, J.F.W. Ed., Plenum Press, 1973 and in Greene, T.W. and Wuts, P.G.M., "Protective Groups in Organic Synthesis", John Wiley & Sons, 1991.

The formation of a desired compound salt is achieved using standard techniques. For example, the neutral compound is treated with an acid or

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base in a suitable solvent and the formed salt is isolated by filtration, extraction or any other suitable method.

The formation of solvates of the compounds of the invention will vary depending on the compound and the solvate. In general, solvates are formed by dissolving the compound in the appropriate solvent and isolating the solvate by cooling or using an antisolvent. The solvate is typically dried or azeotroped under ambient conditions.

Prodrugs of the compounds of the invention may be conventional esters formed with available hydroxy, amino or carboxyl group. For example, when R^1 , R^2 or R^3 is OH in a compound of Formulae I, II and/or III, it may be acylated using an activated acid in the presence of a base, and optionally, in inert solvent (e.g. an acid chloride in pyridine). Some common esters which have been utilized as prodrugs are phenyl esters, aliphatic (C_8 - C_{24}) esters, acyloxymethyl esters, carbamates and amino acid esters.

A radiolabeled compound of the invention may be prepared using standard methods known in the art. For example, tritium may be incorporated into a compound of the invention using standard techniques, for example by hydrogenation of a suitable precursor to a compound of the invention using tritium gas and a catalyst. Alternatively, a compound of the invention containing radioactive iodo may be prepared from the corresponding trialkyltin (suitably trimethyltin) derivative using standard iodination conditions, such as [125] sodium iodide in the presence of chloramine-T in a suitable solvent, such as dimethylformamide. The trialkyltin compound may be prepared from the corresponding non-radioactive halo, suitably iodo, compound using standard palladium-catalyzed stannylation conditions, for example hexamethylditin in the presence of tetrakis(triphenylphosphine) palladium (0) in an inert solvent, such as dioxane, and at elevated temperatures, suitably 50-100°C.

IV. Uses

As hereinbefore mentioned, the inventors have prepared novel compounds of the Formulae I, II and III. Accordingly, the present invention includes all uses of the compounds of the invention including their use in

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therapeutic m thods and compositions for modulating c II proliferation, their use in diagnostic assays and their use as research tools.

In one aspect, the present invention provides a method for modulating cell proliferation comprising administering an effective amount of a compound of the invention to a cell or animal in need thereof. Preferably, the invention provides a method of inhibiting cell proliferation comprising administering an effective amount of a compound of the invention to a cell or animal in need thereof. In particular, the method of the invention is useful in inhibiting the proliferation of abnormal but not normal cells. Abnormal cells include any type of cell that is causative of or involved in a disease or condition and wherein it is desirable to modulate or inhibit the proliferation of the abnormal cell to treat the disease or condition. Examples of abnormal cells include malignant or cancerous cells as well as cell that over-proliferate in inflammatory conditions.

It has been determined that some of the compounds of the invention are very effective at killing cancer cells while at the same time they do not kill normal cells. These properties make the compounds of the invention extremely useful as anti-cancer agents. Accordingly, in one embodiment, the present invention provides a method of inhibiting the proliferation of a cancer cell comprising administering an effective amount of a compound of the invention to a cell or animal in need thereof.

The cancer cell that can be treated with a compound of the invention may be any type of cancer including, but not limited to, hematopoietic malignancies, including leukemias, lymphomas, and myelomas as well as other types of cancer including sarcomas, carcinomas, melanomas, adenomas, nervous system cancers and genitourinary cancers. Examples of leukemias include acute lymphoblastic leukemia (ALL), acute myelocytic leukemia (AML), chronic myeloid leukemia (CML), chronic lymphocytic leukemia (CLL) and juvenile myelo-monocytic leukemia (JMML). The types of ALL that may be treated with the compounds of the invention include cells that express a bcr-abl fusion protein, such as Philadelphia positive ALL cells, as well as Philadelphia negative ALL cells. Examples of lymphomas include B-

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cell Burkitt's lymphoma, Hodgkin's lymphomas, non-Hodgkin's lymphomas, including the Ki-1 positive anaplastic large cell lymphomas, T cell lymphomas and rare lymphomas such as the histiocytic lymphomas. Examples of myelomas include multiple myelomas.

- In a specific embodiment, the present invention provides a method of inhibiting the proliferation of a cancer cell comprising administering an effective amount of a compound selected from the group of compounds:
 - (E,E)-2-(benzylamido)-3-styrylacrylonitrile (CR1);
 - (E,E)-2-(benzylamido)-3-(3,4-dimethoxystyryl)acrylonitrile (CR2);
- 10 (E,E)-2-(benzylamido)-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR3);
 - (E,E)-2-(benzylamido)-3-(3,4-dihydroxystyryl)acrylonitrile (CR4);
 - (E,E)-2-(phenylethylamido)-3-(3,4-dimethoxystyryl)acrylonitrile (CR5);
 - (*E,E*)-2-(phenylethylamido)-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR8);
- (E,E)-2-(phenylpropylamido)-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR9);
 - (E,E)-2-(3,4-dihydroxybenzylamido)-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR11);
 - (E,E)-2-thioacetamido-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR12);
- 20 (E,E)-2-acetamido-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR13);
 - (E,E)-2-carboxy-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR14);
 - (E,E)-2-carbomethoxy-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR15);
 - (E,E)-2-acetamido-3-[3,4-bis(t-butyldimethylsilyloxystyryl)]acrylonitrile(CR16);
 - (E,E)-2-acetamido-3-(3,4-dihydroxystyryl)acrylonitrile (CR17);
- 25 (E,E)-2-(benzylamido)-3-(3,4-bis(t-butyldimethylsilyloxystyryl))acrylonitrile (CR18);
 - (E,E)-2-(3,4 dihydroxybenzylamido)-3-styrylacrylonitrile (CR19);
 - (E,E)-2-(3,4 dihydroxybenzylamido)-3-[3,4-bis(t-
 - butyldimethylsilyloxystyryl)]acrylonitrile (CR20);
- 30 (E,E)-2-(3,4 dihydroxybenzylamido)-3-(3,4-dihydroxystyryl)acrylonitrile (CR21);

(E,E)-2- $(\beta$ -ethanolamido)-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR24);

(E,E)-2-(benzylamido)-3-(4-nitrostyryl)acrylonitrile (CR27);

(E,E)-2-(3,4-dihydroxybenzylamido)-3-(4-nitrostyryl)acrylonitrile(CR28); and

(E,E)-2-(1-amino-2,2-dicyanoethenyl)-3-(4-nitrostyryl)acrylonitrile (CR29).

In a preferred embodiment, the present invention provides a method of inhibiting the proliferation of a cancer cell comprising administering an effective amount of a compound selected from the group of compounds:

(E,E)-2-(benzylamido)-3-(3,4-dihydroxystyryl)acrylonitrile (CR4);

10 (E,E)-2-(3,4-dihydroxybenzylamido)-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR11);

(E,E)-2-acetamido-3-(3,4-dihydroxystyryl)acrylonitrile (CR17);

(E,E)-2-(3,4 dihydroxybenzylamido)-3-styrylacrylonitrile (CR19);

(E,E)-2-(3,4 dihydroxybenzylamido)-3-(3,4-dihydroxystyryl)acrylonitrile

(CR21); and

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(E,E)-2- $(\beta$ -ethanolamido)-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR24).

One skilled in the art can determine which compounds of the invention would have therapeutic utility, for example, in inhibiting cell proliferation in any type of cancer or cell proliferative disorder. Compounds may be examined for their efficacy in inhibiting cell growth in cell proliferation assays such as those described herein in Examples 35-56. Accordingly, the methods, uses and compositions of the invention are meant to include only those compounds having the desired effect.

The ability of the compounds of the Invention to inhibit the growth of cancer cells, in particular hematopoetic cell malignancies, in vitro and in vivo was examined. Several of the compounds tested were found to eliminate cancerous cell growth in culture at sub-micro-molar doses. In particular, CR4, CR11 and CR19 were found to be highly effective against a variety of cell types, such as Acute Lymphoblastic Leukemia, Philadelphia positive Leukemia and Acute Myeloid Leukemia. Low nanomolar doses of both CR4 and CR19 were highly toxic to cancer cells, while normal cell growth and

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differentiation were unaff cted. These effects were obtained by long term exposure to low levels of the compounds. Accordingly, in on aspect, this invention provides a method of inhibiting the proliferation of a hematopoietic cancer cell by administering an effective amount of a compound of the invention, preferably, CR4 or CR11 or CR19, to a cell or animal in need thereof.

It has been determined that the compound CR4 is capable of effectively killing human Philadelphia positive acute lymphoblastic leukemia cells in vivo, using a murine model. CR4 efficiently reduced tumor load and infiltration of the organs by the ALL cells. The doses required to eliminate cancer cell growth do not result in detectable non-specific damage to the animal.

It has also been determined that the compounds of the invention, such as CR4 and CR11, are effective as ex vivo purging agents. For ex vivo administration, bone marrow cells may be removed from a patient with cancer and purged ex vivo with a compound of the invention. Such a purging will kill the tumor cells while leaving the normal bone marrow cells intact. After purging, the cells can be washed and reintroduced into the patient.

During *ex vivo* purging assays the cells were exposed to relatively high doses of the compounds (50μM-100μM) for short (1-24 hours) periods of time, resulting in the elimination of cancer cell growth, while normal bone marrow cells exposed to the same doses over the same period of time were relatively unaffected. Cancer cell death was effected by the induction of apoptosis. Accordingly, in another aspect of the invention, there is provided a method for killing cancer cells by *ex vivo* treatment of bone marrow from a patient with cancer with a compound of the invention, preferably CR4 and CR11 and then re-introducing the treated (or purged) bone marrow into the patient.

In addition to cancer, the compounds of the invention are useful in treating other conditions involving aberrant or abnormal cell proliferation. Other cell proliferative disorders that may be treated by the present invention include inflammatory diseases, allergies, autoimmune disease, graft rejection psoriasis, restenosis, artherosclerosis, and any other disorder wherein it is

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d sirable to inhibit, prevent or suppress cell growth. Compounds of the invention may be tested for the ir efficacy in a particular cell proliferation disorder using assays and techniques known to those of skill in the art. For example, the following references provide assays for various conditions. Rheumatoid Arthritis: "Regulation of IL-15 - Simulated TNF-alpha Production by Rolipram", Journal of Immunology (1999) volume 163 page 8236 by C. S. Kasyapa et al. Allergy: "A novel Lyn-Binding Peptide Inhibitor Blocks Eosinophil Differentiation, Survival, and Airway eosinophilic inflammation". Journal of Immunology (1999) volume 163 page 939 by T. Adachi et al. Psoriasis: Journal of Immunology (2000) volume 165 page 224 "Inhibition of Keratinocyte apoptosis by IL-15: a new paramete in the pathegenosis of psoriasis" by R. Uchert (there is an umlatt over the U). Psoriasis: International Archives of allergy and Immunology (2000) Volume 123 page 275. "T-cell receptor mimic peptides and their potential application in T-cell mediated disease" by A. H. Enk.

The compounds of the invention are tyrosine kinase modulators and are useful in modulating tyrosine kinase activity, including the inhibition of tyrosine kinase activity, for the treatment of various conditions such as all proliferative disorders as mentioned above. Accordingly, the invention provides a method of modulating tyrosine kinase activity by administering an effective amount of a compound of the invention to a cell or animal in need thereof. In a further aspect, the invention provides a method of inhibiting tyrosine kinase activity by administering an effective amount of a compound of the invention to a cell or animal in need thereof.

While the compounds of the invention may act by inhibiting tyrosine kinase activity, one of skill in the art will appreciate that other modes or mechanisms of action for the compounds of the invention are possible.

The compounds of the invention are preferably formulated into pharmaceutical compositions for administration to human subjects in a biologically compatible form suitable for administration in vivo. Accordingly, in another aspect, the present invention provides a pharmaceutical composition

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comprising a compound of the invention in admixture with a suitable diluent or carrier.

The compositions containing the compounds of the invention can be prepared by known methods for the preparation of pharmaceutically acceptable compositions which can be administered to subjects, such that an effective quantity of the active substance is combined in a mixture with a pharmaceutically acceptable vehicle. Suitable vehicles are described, for example, in Remington's Pharmaceutical Sciences (Remington's Pharmaceutical Sciences, Mack Publishing Company, Easton, Pa., USA 1985). On this basis, the compositions include, albeit not exclusively, solutions of the substances in association with one or more pharmaceutically acceptable vehicles or diluents, and contained in buffered solutions with a suitable pH and iso-osmotic with the physiological fluids.

The compounds of this invention may be used in the form of the free base, in the form of salts, solvates and as hydrates. All forms are within the scope of the invention. Acid addition salts may be formed and provide a more convenient form for use; in practice, use of the salt form inherently amounts to use of the base form. The acids which can be used to prepare the acid addition salts include preferably those which produce, when combined with the free base, pharmaceutically acceptable salts, that is, salts whose anions are non-toxic to the animal organism in pharmaceutical doses of the salts, so that the beneficial properties inherent in the free base are not vitiated by side effects ascribable to the anions. Although pharmaceutically acceptable salts of the basic compounds are preferred, all acid addition salts are useful as sources of the free base form even if the particular salt per se is desired only as an intermediate product as, for example, when the salt is formed only for the purposes of purification and identification, or when it is used as an intermediate in preparing a pharmaceutically acceptable salt by ion exchange procedures.

Pharmaceutically acceptable salts within the scope of the invention include those derived from the following acids; mineral acids such as hydrochloric acid, sulfuric acid, phosphoric acid and sulfamic acid; and

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organic acids such as acetic acid, citric acid, lactic acid, tartaric acid, malonic acid, methanesuffonic acid, ethanesulfonic acid, benzenesulfonic acid, p-toluenesulfonic acid, cyclohexylsulfamic acid, quinic acid, and the like.

In accordance with the methods of the invention, the described compounds or salts or solvates thereof may be administered to a patient in a variety of forms depending on the selected route of administration, as will be understood by those skilled in the art. The compositions of the invention may be administered orally or parenterally. Parenteral administration includes intravenous, intraperitoneal, subcutaneous, intramuscular, transepithelial, nasal, intrapulmonary, intrathecal, rectal and topical modes of administration. Parenteral administration may be by continuous infusion over a selected period of time.

A compound of the invention or a salt or solvate thereof may be orally administered, for example, with an inert diluent or with an assimilable edible carder, or it may be enclosed in hard or soft shell gelatin capsules, or it may be compressed into tablets, or it may be incorporated directly with the food of the diet. For oral therapeutic administration, the compound of the invention may be incorporated with excipient and used in the form of ingestible tablets, buccal tablets, troches, capsules, elixirs, suspensions, syrups, wafers, and the like.

A compound of the invention may also be administered parenterally or intraperitoneally. Solutions of a compound of the invention as a free base or pharmacologically acceptable salt or solvate can be prepared in water suitably mixed with a surfactant such as hydroxypropylcellulose. Dispersions can also be prepared in glycerol, liquid polyethylene glycols, DMSO and mixtures thereof with or without alcohol, and in oils. Other solvents include taxol formulation, CMC, Tween20. Under ordinary conditions of storage and use, these preparations contain a preservative to prevent the growth of microorganisms. A person skilled in the art would know how to prepare suitable formulations. Conventional procedures and ingredients for the selection and preparation of suitable formulations are described, for example, in Remington's Pharmaceutical Sciences (1990 - 18th edition) and in The

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United States Pharmacopeia: The National Formulary (USP 24 NF19) published in 1999.

The pharmaceutical forms suitable for Injectable use include sterile aqueous solutions or dispersion and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersions. In all cases the form must be sterile and must be fluid to the extent that easy syringability exists.

The compounds of the invention may be administered to an animal alone or in combination with pharmaceutically acceptable carriers, as noted above, the proportion of which is determined by the solubility and chemical nature of the compound, chosen route of administration and standard pharmaceutical practice.

The dosage of the compounds and/or compositions of the invention can vary depending on many factors such as the pharmacodynamic properties of the compound, the mode of administration, the age, health and weight of the recipient, the nature and extent of the symptoms, the frequency of the treatment and the type of concurrent treatment, if any, and the clearance rate of the compound in the animal to be treated. One of skill in the art can determine the appropriate dosage based on the above factors. The compounds of the invention may be administered initially in a suitable dosage that may be adjusted as required, depending on the clinical response. As an example, the compounds of the invention can be administered in a range from about 1 nanomolar to about 100 micromolar, preferably 50 nanomolar to 50 micromolar. For ex vivo treatment of cells over a short period, for example for 30 minutes to 1 hour or longer, higher doses of compound may be used than for long term in vivo therapy; for example, concentrations of 50µM or higher may be used.

The present invention also includes a use of a compound or composition of the invention in order to inhibit cell proliferation, preferably cancer cell proliferation. The present invention further includes a use of a compound or a composition of the invention to prepare a medicament to inhibit cell proliferation, preferably cancer cell proliferation.

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The compounds of the invention can be used alone or in combination with other agents that modulate tyrosine kinase activity or in combination with other types of treatment (which may or may not modulate tyrosine kinase activity) for cell proliferative disorders. Agents known in the art that inhibit tyrosine kinase activity include, but are not limited to, antisense nucleic acid and ribozymes targeted to nucleic acid encoding a receptor tyrosine kinase, antibodies able to modulate tyrosine kinase activity and other small molecule tyrosine kinase inhibitors such as those described in US 5,891,917, US 5,217,999, US 5,773,476, US 5,935,993, US 5,656,655, US 5,677,329 and US 5,789,427. There are various examples of other types of treatment for cell proliferative disorders currently used to treat different types of cancers. The general treatments are based on the cancer type and do not specifically target tyrosine kinase activity. In a particular aspect of the present invention, the compounds of the invention may be used in combination with other therapies and therapeutics to treat leukemia.

In addition to the above-mentioned therapeutic uses, the compounds of the invention are also useful in diagnostic assays, screening assays and as research tools.

In diagnostic assays the compounds of the invention may be useful in identifying or detecting a cell proliferative disorder. In such an embodiment, the compounds of the invention may be radiolabelled (as hereinbefore described) and contacted with a population of cells. The presence of the radiolabelled on the cells may indicate a cell proliferative disorder. In a specific embodiment, the radiolabelled compounds of the invention may be used to detect the presence of cells expressing a bcr-abl fusion protein.

In screening assays, the compounds of the invention may be used to identify other compounds that modulate cell proliferation or tyrosine kinase activity. As research tools, the compounds of the invention may be used in receptor binding assays and assays to study the localization of tyrosine kinases. In such assays, the compounds may also be radiolabelled.

Novel compounds and methods for modulating cell proliferation also are described in PCT/CA 01/____, filed April 12, 2001, which claims the

priority benefit of U.S. Provisional Application Serial No. 60/196,936, filed April13, 2000, the disclosur s of which are incorporated herein by reference in their entirety.

The following non-limiting examples are illustrative of the present invention:

EXAMPLES

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Materials and Methods For Examples 1-34

¹H NMR spectra were obtained on a Varian Unity Plus spectrometer (USA) at 500 MHz with tetramethylsilane (TMS, Me₄Si) as an internal standard (δ =0). Electrospray mass spectra were recorded on an API III Plus triple quadrupole mass spectrometer (USA), with a direct introduction of the samples into the ionization source. Thin layer chromatography was performed with UV-254 aluminum-backed TLC sheets of 0.25 mm thickness (Kieselgel 60 F₂₅₄, Merck, Germany). HPLC separation of the compound of Example 13 was performed on a Waters 600 chromatograph (USA), column Nova-Pak C18 3.9 \times 300 mm (Waters, USA). Vacuum distillations were done using Kugelrohr apparatus (Aldrich, USA) at stated temperatures of an oven. 3,5-Dimethoxy-4-hydroxycinnamaldehyde, 4-nitrocinnamaldehyde, 3,4acid, 3,4-dihydroxycinnamic dimethoxycinnamic dimethoxybenzylamine, benzylamine, phenylethylamine, phenylpropylamine, methyl cyanoacetate, 2-cyanothioacetamide, 2-cyanoacetamide, cyanoacetic acid, β-ethanolamine, 2-amino-1-propene-1,1,3-tricarbonitrile were purchased from Aldrich (USA) and were used as received. The reagents were from Aldrich (USA), Solvents were purchased from Caledon (Canada).

Example 1: N-(Cyanoacetyl)3,4-dimethoxybenzylamide (A₁)

To 3,4-dimethoxybenzylamine (2.7 ml, 18 mmol) methyl cyanoacetate was added (1.6 ml, 18 mmol). The reaction was heated for 14 h at 100°C.

Cooling gave a dark brown solid which was recrystallized from ethanol to give 2.90 g of th product (69% yield).

The product gave the following analytical data:

NMR (CD₃COCD₃, δ , ppm): 3.62 (s, 2H, CH₂CN), 3.78 (s, 6H, (OMe)₂), 4.34 (br.s., 2H, NHCH₂Ph), 6.84 (dd, 1H, J 1.95 and 8.1 Hz, H⁵), 6.88 (d, 1H, J 8.1 Hz, H⁵), 6.93 (d, 1H, J 1.95 Hz, H²), 7.80 (br.s., 1H, NH).

MS, m/e (rel. intensity, %): 235 (19) $[M+H]^{+}$, 252 (100) $[M+NH_{4}]^{+}$, 257 (33) $[M+Na]^{+}$.

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Example 2: N-(Cyanoacetyl)3,4-dihydroxybenzylamide (A2)

To N-(cyanoacetyl)3,4-dimethoxybenzylamide (Example 1, 0.2 g, 0.85 mmol) in 20 ml of CH_2Cl_2 boron tribromide was added under argon at -78°C (0.24 ml, 2.56 mmol) in 2.5 ml of CH_2Cl_2 . After 2 h the reaction was brought to room temperature and stirred overnight. The reaction was cooled to 0°C, 10 ml of 1N HCl was added, the solution was extracted with 3 × 50 ml of ethyl acetate, the organic phase was washed to neutral pH, dried with MgSO₄, and taken to dryness. The residue was purified by silica gel chromatography (CHCl₃-MeOH, 20:1) to give a yellow solid (0.07 g, 40% yield). The product gave the following analytical data:

NMR (CD₃COCD₃, δ , ppm): 2.83 (s, (OH)₂), 3.60 (s, 2H, CH₂CN), 4.25 (br.s., 2H, NHCH₂Ph), 6.63 (dd, 1H, J 1.95 and 8.1 Hz, H⁶), 6.75 (d, 1H, J 8.1 Hz, H⁵), 6.79 (d, 1H, J 1.95 Hz, H²), 7.71 (br.s., 1H, NH).

25 MS, m/e (rel. intensity, %): 207 (38) $[M+H]^+$, 224 (100) $[M+NH_4]^+$, 229 (2.6) $[M+Na]^+$.

Example 3: (E,E)-2-(3,4-Dihydroxybenzylamido)-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR11)

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To 3,5-dimethoxy-4-hydroxycinnamaldehyde (0.042 g, 0.2 mmol) and N-(cyanoacetyl)3,4-dihydroxybenzylamide (Example 2, 0.042 g, 0.2 mmol) in 10 ml of ethanol 3 mg of β -alanine was added and the reaction was refluxed for 6 h. Water was added and the solid was recrystallized from 5 ml of ethanol twice to give 0.06 g (75%) of a red solid. The product gave the following analytical data:

NMR (CD₃COCD₃, δ , ppm): 2.81 (s, (OH)₃), 3.89 (s, 6H, (OMe)₂), 4.39 (br.s., 2H, NHCH₂Ph), 6.68 (dd, 1H, J 1.95 and 8.1 Hz, H⁶), 6.76 (d, 1H, J 8.1 Hz, H⁵), 6.86 (d, 1H, J 1.95 Hz, H²), 7.07 (br.s., 2H, H²⁺⁶), 7.16 (dd, 1H, J 11.7 and 15.1 Hz, PhCCHCCN olefinic), 7.37 (d, 1H, J 15.1 Hz, PhCH olefinic), 7.70 (br.s., 1H, NH), 7.98 (dd, 1H, J 0.75 and 11.7 Hz, CHCN olefinic). MS, m/e (rel. intensity, %): 397 (100) [M+H]⁺, 414 (14) [M+NH₄]⁺.

15 Example 4: N-(Cyanoacetyl)benzylamide (A₃)

The compound was prepared as described in Example 1 by adding methyl cyanoacetate (1.3 ml, 14 mmol) to benzylamine (1.5 ml, 14 mmol). The compound was distilled *in vacuo* directly from the reaction mixture (Kugelrohr apparatus (Aldrich), 0.1 mm Hg, T. oven 180-190°C) to give an off-white solid (2.34 g, 95%). The product gave the following analytical data:

NMR (CD₃COCD₃, δ , ppm): 3.39 (s, 2H, CNCH₂), 4.46 (d, 2H, J 5.4 Hz, NHCH₂Ph), 6.40 (br.s., 1H, NH), 7.24-7.36 (m, 5H, Ph).

MS, m/e (rel. intensity, %): 175 (64) [M+H]⁺, 192 [M+NH₄]⁺.

Example 5: 3,4-Dimethoxycinnamyl alcohol (A₆)

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To a solution of 0.42 g (2.0 mmol) of 3,4-dimethoxycinnamic acid in 50 ml MeOH was added SOCl₂ (50 µl) and the mixture was stirred at 60°C for 5 h. Methanol was taken to dryness and the obtained 3,4-dimethoxycinnamic acid methyl ester was reduced with 1M THF solution of diisobutylaluminum hydride (8.0 mmol) in absolute THF (50 ml) at 20°C for 1 h. Water was added, the mixture was extracted with EtOAc, dried with MgSO₄ and distilled *in vacuo* (Kugelrohr apparatus (Aldrich), 0.1 mm Hg, T. oven 185-190°C) giving an offwhite solid, yield 0.36 g (92%), m.p. 70-71°C. The product gave the following analytical data:

NMR (CD₃COCD₃, δ , ppm): 3.77, 3.82 (2 × s, 2 × 3H, OMe + OMe), 4.19 (d, 2H, J 5.0 Hz, CH₂OH), 6.25 (dt, 1H, J 5.0 and 15.5 Hz, PhCCH olefinic), 6.51 (d, 1H, J 15.5 Hz, PhCH olefinic), 6.89 (m, 2H, H⁵⁺⁶), 7.05 (br.s., 1H, H²). MS, m/e (rel. intensity, %): 177 (100) [M-OH]+, 195 (4) [M+H]⁺, 212 (59) [M+NH₄]⁺, 217 (26) [M+Na]⁺.

Example 6: 3,4-Dimethoxycinnamaldehyde (A7)

To a mixture of pyridinium dichromate (3.88 g, 10.3 mmol) and 4 g of finely grounded freshly activated molecular sieves 3Å in 20 ml of CH₂Cl₂ 3,4-dimethoxycinnamyl alcohol in 10 ml of CH₂Cl₂ (Example 5, 1.00 g, 5.1 mmol) was added. The reaction was stirred for 2 h, 0.5 ml of methanol was added, the residue was passed through silica gel and washed with 300 ml of ethyl acetate. After evaporation the compound was purified by silica gel chromatography (hexane-EtOAc, 5:1) leading to a crystallizing oil (0.62 g, 63%).

The product gave the following analytical data:

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NMR (CD₃COCD₃, δ , ppm): 3.90 (2 × s, 2 × 3H, OCH₃ + OCH₃), 6.70 (dd, 1H, J 7.6 and 16.0 Hz, PhC=CH olefinic), 7.05 (d, 1H, J 8.3 Hz, H⁵), 7.28 (dd, 1H, J 1.4 and 8.3 Hz, H⁶), 7.37 (d, 1H, J 1.4 Hz, H²), 7.60 (d, 1H, J 16.0 Hz, PhCH olefinic), 9.65 (d, 1H, J 7.6 Hz, CHO).

5 MS, m/e (rel. intensity, %): 193 (100) [M+H]⁺, 210 (26) [M+NH₄]⁺.

Example 7: (E,E)-2-(Benzylamido)-3-(3,4-dimethoxystyryl) acrylonitrile (CR2)

The compound was prepared as described in Example 3, by adding 3,4-dimethoxycinnamaldehyde (Example 6, 0.04 g, 0.2 mmol) to N-(cyanoacetyl)benzylamide (Example 4, 0.036 g, 0.2 mmol). After refluxing for 1 h and recrystallization from ethanol a yellow solid was obtained (0.045 g, 62%). The product gave the following analytical data:

NMR (CD₃COCD₃, δ, ppm): 3.90 (s, 2 × 3H, OMe + OMe), 4.57 (d, 2H, J < 2 Hz, NHCH₂Ph), 7.08 (br.s., 1H, H²), 7.17 (dd, 1H, J 11.5 and 15.2 Hz, PhCCHCCN olefinic), 7.23-7.42 (m, 8H, aromatic + H⁵ + H⁶ + PhCH olefinic), 7.90 (br.t, 1H, NH), 8.05 (dd, 1H, J 0.55 and 11.5 Hz, CHCN olefinic).

20 Example 8: (E,E)-2-(Benzylamido)-3-(3,4-dihydroxystyryl)acrylonitrile (CR4) – Method A

Boron tribromide (0.033 ml, 0.34 mmol) was added to (E,E)-2-(benzylamido)-3-(3,4-dimethoxystyryl)acrylonitrile (Example 7, 0.04 g, 0.11 mmol). The residue was purified by silica gel chromatography (CHCl₃-MeOH, 10:1) to give an orange solid (0.02 g, 55% yield). The product gave the following analytical data:

NMR (CD₃COCD₃, δ , ppm): 2.86 (br.s., 2H, (OH)₂), 4.55 (m, 2H, NHCH₂Ph), 6.90-7.42 (m, 10H, Ph + Ph' + olefinic), 7.87 (br.s., 1H, NH), 8.02 (dd, 1H, J < 0.5 and 11.4 Hz, CHCN olefinic).

MS, m/e (rel. intensity, %): 295 (61) [M+H-CN]⁺, 321 (100) [M+H]⁺, 338 (30) [M+NH₄]⁺.

Example 9: Methyl ester of 3,4-bis(t-butyldimethylsilyloxy)cinnamic acid (A₈)

To a solution of 3.6 g (20 mmol) of 3,4-dihydroxycinnamic acid in 300 ml MeOH was added SOCl₂ (100 μl) and the mixture was stirred at 60°C for 5 h. Methanol was taken to dryness and the obtained methyl ester was treated up with 10.2 g (68 mmol) of *t*-BuMe₂SiCl and 9.2 g (136 mmol) of imidazole in 100 ml DMF at 50°C for 0.5 h. Mixture was diluted with water and extracted with hexane. Hexane was taken to dryness. The residue was distilled *in vacuo* (Kugelrohr apparatus (Aldrich), 0.1 mm Hg, T. oven 200-210°C) and crystallized from hexane at -20°C giving a white solid, yield 7.5 g (89%), m.p. 57-58°C. The product gave the following analytical data:

Example 10: 3,4-Bis(t-butyldimethylsilyloxy)cinnamyl alcohol (A9)

MS, m/e (rel. intensity, %): 423 (100) $[M+H]^{+}$, 440 (98) $[M+NH_{4}]^{+}$.

The compound was prepared as described in Example 5 by treating of 3,4-dihydroxycinnamic acid bis(BDMS) ether methyl ester (Example 9, 0.42 g, 1.0 mmol) with 1M THF solution of diisobutylaluminum hydride (4.0 mmol) in absolute THF (25 ml) at 20°C for 1 h. After distilling *in vacuo* (Kugelrohr apparatus (Aldrich), 0.1 mm Hg, T. oven 185-200°C) a white viscous oil was obtained, yield 0.33 g (85%). The product gave the following analytical data:

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NMR (CD₃COCD₃, δ , ppm): 0.23, 0.24 (2 × s, 2 × 6H, Me₂Si + Me₂Si), 1.00, 1.02 (2 × s, 2 × 9H, *t*-BuSi) + *t*-BuSi), 4.19 (d, 2H, J 4.9 Hz, CH₂OH), 6.22 (dt, 1H, J 4.9 and 16.0 Hz, PhCCH olefinic), 6.49 (d, 1H, J 16.0 Hz, PhCH olefinic), 6.85 (d, 1H, J 8.2 Hz, H⁵), 6.92 (dd, 1H, J 2.1 and 8.2 Hz, H⁶), 6.97 (d, 1H, J 2.1 Hz, H²).

MS, m/e (rel. intensity, %): 377 (100) [M-OH]⁺, 395 (2) [M+H]⁺, 412 (15) [M+NH₄]⁺.

Example 11: 3,4-Bis(t-butyldimethylsilyloxy)cinnamaldehyde (A₁₀)

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The compound was prepared as described in Example 6 by adding 3,4-bis(t-butyldimethylsilyloxy)cinnamyl alcohol (Example 10, 0.2 g, 0.5 mmol) in 5 ml of CH₂Cl₂ to a mixture of pyridinium dichromate (0.38 g, 1 mmol) and 1 g molecular sieves 3Å in 20 ml of CH₂Cl₂. The residue was passed through silica gel and washed with 300 ml of EtOAc-hexane, 1:1. After evaporation the compound was purified by silica gel chromatography (hexane-EtOAc, 5:1) leading to an oil (0.15 g, 76%). The product gave the following analytical data: NMR (CD₃COCD₃, δ , ppm): 0.26 and 0.28 (2 × s, 2 × 6H, Me₂Si + Me₂Si), 1.01 and 1.02 (2 × s, 2 × 9H, t-BuSi + t-BuSi), 6.60 (dd, 1H, J 7.7 and 15.9 Hz, PhCCH olefinic), 7.01 (dd, 1H, J < 0.5 and 8.9 Hz, H⁶), 7.27 (m, 2H, H²⁺⁵), 7.60 (d, 1H, J 15.9 Hz, PhCH olefinic), 9.65 (d, 1H, J 7.7 Hz, CHO). MS, m/e (rel. intensity, %): 367 (3) [M+H-CN]⁺, 393 (100) [M+H]⁺, 410 (10) [M+NH₄]⁺.

25 Example 12: (E,E)-2-(Benzylamido)-3-(3,4-bis(t-butyldimethylsilyloxystyryl))acrylonitrile (CR18)

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The compound was prepared as described in Exampl 3 by adding 3,4-bis(*t*-butyldimethylsilyloxy)cinnamaldehyde (Example 11, 0.100 g, 0.26 mmol) to N-(cyanoacetyl)benzylamide (Example 4, 0.044 g, 0.26 mmol. After refluxing for 2.5 h purification by silica gel chromatography (hexane-EtOAc, 15:1) provided a yellow solid (0.090 g, 64%). The product gave the following analytical data:

NMR (CD₃COCD₃, δ , ppm): 0.24 and 0.25 (2 × s, 2 × 6H, Me₂Si + Me₂Si), 1.01 and 1.02 (2 × s, 2 × 9H, *t*-BuSi + *t*-BuSi), 4.55 (br.s., 2H, NHCH₂Ph), 7.00 (d, 1H, J 8.5 Hz, H⁴), 7.12 (dd, 1H, J 11.7 and 15.6 Hz, PhCCHCCN olefinic), 7.24-7.43 (m, 8H, aromatic and olefinic protons), 7.93 (br.s., 1H, NH), 8.02 (dd, 1H, J < 0.5 and 11.7 Hz, CHCN olefinic).

MS, m/e (rel. intensity, %): 523 (30) [M+H-CN]⁺, 540 (24) [M+NH₄-CN]⁺, 549 (89) [M+H]⁺, 566 (100) [M+NH₄]⁺.

Example 13: (E,E)-2-(Benzylamido)-3-(3,4-dihydroxystyryl)acrylonitrile (CR4) – Method B

(E,E)-2-Benzylamido-3-[3,4-bis(t-butyldimethylsilyloxystyryl)]

acrylonitrile (Example 12, 0.028 g, 0.052 mmol) was treated with 60 μl of a 1M THF solution of tetra-*n*-butylammonium fluoride in 2 ml of dry THF for 0.5 h at 20°C. After evaporation the compound was dissolved in 5 ml of chloroform-methanol, 20:1, passed through silica gel and washed with chloroform-methanol, 20:1. The residue was purified by HPLC chromatography (MeCN-H₂O₁ 60:40, UV detection at 340 nm) leading to an orange solid (0.010 g, 62%). The analytical data were identical to the compound prepared as described in Example 8.

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Exampl 14: (E,E)-2-(3,4 Dihydroxybenzylamido)-3-styrylacrylonitrile (CR19)

The compound was prepared as described in Example 3 by adding cinnamaldehyde (0.018 ml, 0.14 mmol) to N-(cyanoacetyl)3,4-dihydroxybenzylamide (Example 2, 0.03 g, 0.14 mmol). After refluxing for 2 h and recrystallization from ethanol, a yellow solid was obtained (0.027 g, 59%). The product gave the following analytical data:

NMR (CD₃COCD₃, δ , ppm): 2.82 (br.s., 2H, (OH)₂), 4.39 (br.s., 2H, NHCH₂Ph), 6.70 (dd, 1H, J 1.9 and 8.2 Hz, H⁶), 6.76 (d, 1H, J 8.2 Hz, H⁵), 6.87 (d, 1H, J 1.9 Hz, H²), 7.30 (dd, 1H, J 11.3 and 15.7 Hz, PhCCHCCN olefinic), 7.47 and 7.73 (2 × m, 6H, aromatic protons and PhCH olefinic), 7.82 (br.s., 1H, NH), 8.04 (dd, 1H, J < 0.5 and 11.3 Hz, CHCN olefinic). MS, m/e (rel. intensity, %):321 (100) [M+H]⁺, 338 (65) [M+NH₄]⁺.

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Example 15: (E,E)-2-(3,4 Dihydroxybenzylamido)-3-[3,4-bis(t-butyldimethylsilyloxystyryl)]acrylonitrile (CR20)

The compound was prepared as described in Example 3 by adding 3,4-bis(t-butyldimethylsilyloxy)cinnamaldehyde (Example 11, 0.015 g, 0.038 mmol) to N-(cyanoacetyl)3,4-dihydroxybenzylamide (Example 2, 0.0079 g, 0.038 mmol). After refluxing for 2 h and recrystallization from ethanol a yellow solid was obtained (0.014 g, 64%). The product gave the following analytical data:

NMR (CD₃COCD₃, δ, ppm): 0.22 and 0.24 (2 × s, 2 × 6H, Me₂Si + Me₂Si), 1.01 and 1.03 (2 × s, 2 × 9H, t-BuSi + t-BuSi), 2.72 (br.s., 2H, (OH)₂), 4.41

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(br.s., 2H, NHCH₂Ph), 6.68-7.42 (m, 8H, aromatic and of finic protons), 7.75 (br.s., 1H, NH), 8.00 (dd, 1H, J < 0.5 and 12.0 Hz, CHCN olefinic).

MS, m/e (rel. intensity, %): 555 (5) [M+H-CN]⁺, 572 (8) [M+NH₄-CN]⁺, 581 (46) [M+H]⁺, 598 (100) [M+NH₄]⁺.

Example 16: (E,E)-2-(3,4 Dihydroxybenzylamido)-3-(3,4-dihydroxystyryl)acrylonitrile (CR21)

(E,E)-2-(3,4-Dihydroxybenzylamido)-3-[3,4-bis(t-butyldimethyl-

silyloxystyryl)]acrylonitrile (Example 15, 0.026 g, 0.044 mmol) was treated with 60 µl of a 1M THF solution of tetra-n-butylammonium fluoride in 1.5 ml of dry THF for 0.5 h at 20°C as described in Example 13. After purification, a yellow solid was obtained (0.006 g, 43%). The product gave the following analytical data:

15 NMR (CD₃COCD₃, δ, ppm): 4.38 (s, 1H, NHCH₂Ph), 6.67-7.22 (m, 6H, Ph + Ph'), 7.05 (dd, 1H, J 11.8 and 15.5 Hz, PhC=CH olefinic), 7.34 (d, 1H, J 15.5 Hz, PhCH olefinic), 7.70 (br.s, 1H, NH), 8.00 (d, 1H, J 11.8 Hz, CH=CCN olefinic).

MS, m/e (rel. intensity, %): 186 (86) $[(HO)_2C_6H_3CH=CHCH=CCN]^{\dagger}$, 202 (28), 242 (100) $[M+H-C_6H_3(OH)_2]^{\dagger}$, 353 (13) $[M+H]^{\dagger}$, 370 (6) $[M+NH_4]^{\dagger}$.

Example 17: (E,E)-2-(Benzylamido)-3-styrylacrylonitrile (CR1)

The compound was prepared as described in Example 3 by adding cinnamaldehyde (0.048 ml, 0.38 mmol) to N-(cyanoacetyl)benzylamide (Example 4, 0.066 g, 0.38 mmol). After refluxing for 1 h and recrystallization

from ethanol a white solid was obtained (0.074 g, 68%). The product gave the following analytical data:

NMR (CD₃COCD₃, δ, ppm): 4.55 (s, 1H, NHCH₂Ph), 7.24-7.51 (m, 11H, Ph + Ph' + PhCCHCCN olefinic), 7.72 (br.d, 1H, J 6.5 Hz, CHCN olefinic), 7.98 (br.s., 1H, NH), 8.05 (d, 1H, J 11.7 Hz, PhCH olefinic).

MS, m/e (rel. intensity, %): 289 (100) [M+H]⁺, 306 (92) [M+NH₄]⁺.

Example 18: (E,E)-2-(Benzylamido)-3-(3,5-dimethoxy-4-hydroxystyryl) acrylonitrile (CR3)

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The compound was prepared as described in Example 3 by adding 3,5-dimethoxy-4-hydroxycinnamaldehyde (0.10 g, 0.48 mmol) to N-(cyanoacetyl)benzylamide (Example 4, 0.084 g, 0.48 mmol). After refluxing for 3 h and recrystallization from ethanol, a yellow solid was obtained (0.10 g, 57%). The product gave the following analytical data:

NMR (CD₃COCD₃, δ , ppm): 3.90 (s, 6H, (OMe)₂), 4.55 (m, 2H, NHCH₂Ph), 7.08 (br.s, 2H, H²⁺⁶), 7.17 (dd, 1H, J 11.5 and 15.2 Hz, PhCCHCCN olefinic), 7.22-7.41 (m, 6H, Ph' + PhCH olefinic), 7.90 (br.s., 1H, NH), 8.01 (dd, 1H, J 0.55 and 11.7 Hz, CHCN olefinic).

20 MS, m/e (rel. intensity, %): 275 (14) [M+H-CN-MeOH-MeOH][†], 307 (9) [M+H-CN-MeOH][†], 339 (4) [M+H-CN][†], 365 (100) [M+H][†], 382 (16) [M+NH₄][†].

Example 19: N-(Cyanoacetyl)phenylpropylamide (A₅)

The compound was prepared as described in Example 1 by adding methyl cyanoacetate (0.98 ml, 11.1 mmol) to phenylpropylamine (1.58 ml, 11.1 mmol). The compound was distilled *in vacuo* directly from the reaction

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mixture (Kugelrohr apparatus (Aldrich), 0.1 mm Hg, T. oven 195-200°C) to give an off-white solid (2.18 g, 97%). The product gave the following analytical data:

NMR (CD₃COCD₃, δ, ppm): 1.88 (q, 2H, J 7.3 Hz, PhCCH₂), 2.66 (t, 2H, J 7.3 Hz, PhCH₂), 3.28 (s, 2H, CNCH₂), 3.33 (dt, 2H, J 7.3 and 6.6 Hz, PhCCCH₂), 6.02 (br.s., 1H, NH), 7.15-7.30 (m, 5H, Ph).

MS, m/e (rel. intensity, %): 203 (88) [M+H][†], 220 (100) [M+NH₄][†].

Example 20: N-(Cyanoacetyl)phenylethylamide (A4)

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The compound was prepared as described in Example 1 by adding methyl cyanoacetate (1.1 ml, 12.4 mmol) to phenylethylamine (1.55 ml, 12.4 mmol). The compound was distilled *in vacuo* directly from the reaction mixture (Kugelrohr apparatus (Aldrich), 0.1 mm Hg, T. oven 190-195°C) to give an off-white solid (2.14 g, 91%). The product gave the following analytical data:

NMR (CD₃COCD₃, δ, ppm): 2.80 (t, 2H, J 7.6 Hz, PhCH₂), 3.46 (br.t, 2H, J 7.6 Hz, PhCCH₂), 3.54 (s, 2H, CNCH₂), 7.20-7.31 (m, 5H, Ph), 7.51 (br.s., 1H, NH).

MS, m/e (rel. intensity, %): 189 (100) [M+H]⁺, 206 (99) [M+NH₄]⁺.

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Example 21: (E,E)-2-(Phenylethylamido)-3-(3,4-dimethoxystyryl) acrylonitrile (CR5)

The compound was prepared as described in Example 3 by adding 3,4-dimethoxycinnamaldehyde (Example 6, 0.1 g, 0.52 mmol) to N-(cyanoacetyl)phenylethylamide (Example 20, 0.1 g, 0.52 mmol). After

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refluxing for 1 h and recrystallization from ethanol a yellow solid was obtained (0.12 g, 63%). The product gave the following analytical data:

NMR (CD₃COCD₃, δ , ppm): 2.91 (t, 2H, J 7.5 Hz, Ph'CH), 3.59 (br.t, 2H, J 7.5 Hz, Ph'CCH), 3.88, 3.89 (2 × s, 2 × 3H, OCH₃ + OCH₃), 7.04 (d, 1H, J 8.6 Hz, H5), 7.16 (dd, 1H, J 11.8 and 15.0 Hz, PhC=CH olefinic), 7.20-7.42 (m, 9H, aromatic + olefinic), 7.97 (d, 1H, J 11.8 Hz, CH=CCN olefinic). MS, m/e (ref. intensity, %): 363 (100) [M+H]⁺, 380 (34) [M+NH₄]⁺.

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Example 22: (E,E)-2-(Phenylethylamido)-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR8)

The compound was prepared as described in Example 3 by adding 3,5-dimethoxy-4-hydroxycinnamaldehyde (0.1g, 0.48 mmol) to N-(cyanoacetyl)phenylethylamide (Example 20, 0.091g, 0.48 mmol). The residue was purified by silica gel chromatography (CHCl₃-hexane, 1:1) to give a yellow solid (0.15 g, 83% yield). The product gave the following analytical data:

NMR (CD₃COCD₃, δ, ppm): 2.95 (t, 2H, J 7.6 Hz, CH₂Ph'), 3.62 (m, 2H, CH₂CPh'), 3.94 (s, 6H, (OMe)₂), 7.11 (s, 2H, H²⁺⁶), 7.19 (dd, 1H, J 11.7 and 15.3 Hz, PhCCHCCN olefinic), 7.23-7.36 (m, 5H, Ph'), 7.41 (d, 1H, J 15.3 Hz, PhCH olefinic), 7.45 (br.s., 1H, NH), 7.99 (d, 1H, J 11.7 Hz, CHCN olefinic). MS, m/e (rel. intensity, %): 379 (100) [M+H]⁺, 396 (7) [M+NH₄]⁺.

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Example 23: (E,E)-2-(Phenylpropylamido)-3-(3,5-dimethoxy-4-hydroxystyryl)acrylonitrile (CR9)

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The compound was prepared as described in Example 3 by adding 3,5-dimethoxy-4 hydroxycinnamaldehyde (0.10 g, 0.48 mmol) to N-(cyanoacetyl)phenylpropylamide (Example 19, 0.097 g, 0.48 mmol). After refluxing for 3 h the residue was purified by silica gel chromatography (CHCl₃-hexane, 1:1) to give a brown solid (0.17 g, 90% yield). The product gave the following analytical data:

NMR (CD₃COCD₃, δ, ppm): 2.09 (q, 2H, J 7.5 Hz, NHCCH₂CPh³), 2.85 (t, 2H, J 7.5 Hz, CH₂Ph³), 3.57 (m, 2H, CH₂CPh³), 4.06 (s, 6H, (OMe)₂), 7.24 (s, 2H, H²⁺⁶), 7.32 (dd, 1H, J 11.7 and 15.3 Hz, PhCCHCCN olefinic), 7.33-7.46 (m, 5H, Ph³), 7.53 (d, 1H, J 15.3 Hz, PhCH olefinic), 7.58 (br.s., 1H, NH), 8.11 (d, 1H, J 11.7 Hz, CHCN olefinic).

MS, m/e (rel. intensity, %): 331 (40), 348 (30), 359 (34), 376 (32), 393 (100) [M+H]⁺, 410 (24) [M+NH₄]⁺.

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Example 24: (E,E)-2-Thioacetamido-3-(3,5-dimethoxy-4-hydroxystyryl) acrylonitrile (CR12)

The compound was prepared as described in Example 3 by adding 3,5-dimethoxy-4-hydroxycinnamaldehyde (0.15 g, 0.72 mmol) to 2-cyanothioacetamide (0.073 g, 0.72 mmol). After refluxing for 1 h the residue was purified on a TLC-plate in hexane-ethyl acetate, 1:1 to give a red solid (0.10 g, 52%). The product gave the following analytical data:

NMR (CD₃COCD₃, δ , ppm): 2.85 (br.s., OH + NH₂), 3.91 (s, 6H, (OMe)₂), 7.11 (s, 2H, H²⁺⁶), 7.20 (dd, 1H, J 11.6 and 15.1 Hz, PhCCHCCN olefinic), 7.46 (d,

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1H, J 15.1 Hz, PhCH olefinic), 8.22 (dd, 1H, J 0.73 and 11.6 Hz, CHCN olefinic).

MS, m/e (rel. intensity, %): 289 (100), 291 (60) [M+H]⁺, 312 (8) [M+Na]⁺.

Example 25: (E,E)-2-Acetamido-3-(3,5-dimethoxy-4-hydroxystyryl) acrylonitrile (CR13)

The compound was prepared as described in Example 3 by adding 3,5-dimethoxy-4-hydroxycinnamaldehyde (0.1 g, 0.48 mmol) to 2-cyanoacetamide (0.04 g, 0.48 mmol). After refluxing for 3 h and recrystallization from ethanol an orange solid was obtained (0.083 g, 63%). The product gave the following analytical data:

NMR (CD₃COCD₃, δ , ppm): 2.82-2.88 (br.s., OH + NH₂), 3.90 (s, 6H, (OMe)₂), 7.08 (s, 2H, H²⁺⁶), 7.16 (dd, 1H, J 11.6 and 15.1 Hz, PhCCHCCN olefinic), 7.38 (d, 1H, J 15.1 Hz, PhCH olefinic), 7.96 (dd, 1H, J 0.73 and 11.6 Hz, CHCN olefinic).

MS, m/e (rel. intensity, %): 275 (100) [M+H]⁺, 292 (28) [M+NH₄]⁺.

Example 26: (E,E)-2-Carboxy-3-(3,5-dimethoxy-4-hydroxystyryl) acrylonitrile (CR14)

The compound was prepared as described in Example 3 by adding 3,5-dimethoxy-4-hydroxycinnamaldehyde (0.15 g, 0.72 mmol) to cyanoacetic acid (0.061 g, 0.72 mmol). After refluxing for 1 h and recrystallization from ethanol a yellow solid was obtained (0.15 g, 75%). The product gave the following analytical data:

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NMR (CD₃COCD₃, δ, ppm): 3.00 (br.s., OH), 3.91 (s, 6H, (OMe)₂), 7.12 (s, 2H, H²⁺⁶), 7.21 (dd, 1H, J 11.6 and 15.1 Hz, PhCCHCCN olefinic), 7.50 (d, 1H, J 15.1 Hz, PhCH olefinic), 8.04 (dd, 1H, J 0.73 and 11.6 Hz, CHCN olefinic).

5 MS, m/e (rel. intensity, %): 276 (66) [M+H]⁺, 293 (100) [M+NH₄]⁺.

Example 27: (E,E)-2-Carbomethoxy-3-(3,5-dimethoxy-4-hydroxystyryl) acrylonitrile (CR15)

The compound was prepared as described in Example 3 by adding 3,5-dimethoxy-4-hydroxycinnamaldehyde (0.15 g, 0.72 mmol) to methyl cyanoacetate (0.064 ml, 0.72 mmol). After refluxing for 1 h and recrystallization from ethanol an orange solid was obtained (0.2 g, 90%). The product gave the following analytical data:

NMR (CD₃COCD₃, δ, ppm): 2.84 (br.s., OH), 3.84 (s, 3H, COOMe), 3.91 (s, 6H, (OMe)₂), 7.12 (s, 2H, H²⁺⁶), 7.21 (dd, 1H, J 11.6 and 15.1 Hz, PhCCHCCN olefinic), 7.53 (d, 1H, J 15.1 Hz, PhCH olefinic), 8.05 (dd, 1H, J 0.73 and 11.6 Hz, CHCN olefinic).

MS, m/e (rel. intensity, %): 290 (100) [M+H]⁺, 307 (99) [M+NH₄]⁺.

Example 28: (E,E)-2-Acetamido-3-[3,4-bis(t-butyldimethylsilyloxystyryl)]acrylonitrile(CR16)

The compound was prepared as described in Example 3 by adding 3,4-bis(t-butyldimethylsilyloxy)cinnamaldehyde (Example 11, 0.15 g, 0.38 mmol) to 2-cyanoacetamide (0.032 g, 0.38 mmol). After refluxing for 0.5 h,

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purification by silica gel chromatography (hexan -EtOAc, 5:1) provided a crystallizing oil (0.10 g, 57%).

NMR (CD₃COCD₃, δ , ppm): 0.22 and 0.24 (2 × s, 2 × 6H, Me₂Si + Me₂Si), 1.01 and 1.03 (2 × s, 2 × 9H, *t*-BuSi + *t*-BuSi), 7.01, 7.23-7.29 (m, 3H, aromatic), 7.11 (dd, 1H, J 11.9 and 15.3 Hz, PhC=CH olefinic), 7.40 (d, 1H, J 15.3 Hz, PhCH olefinic), 7.98 (d, 1H, J 11.9 Hz, CH=CCN olefinic). MS, m/e (rel. intensity, %): 459 (100) [M+H]⁺, 476 (89) [M+NH₄]⁺.

Example 29: (E,E)-2-Acetamido-3-(3,4-dihydroxystyryl)acrylonitrile 10 (CR17)

(E,E)-2-Acetamido-3-(3,4-bis(t-butyldimethylsilyloxystyryl)) acrylonitrile (Example 28, 0.1 g, 0.22 mmol) was treated with an excess of a 1M THF solution of tetra-*n*-butylammonium fluoride in benzene for 0.5 h at 20°C as described in Example 13. After purification, a yellow solid was obtained (0.04 g, 85%). The product gave the following analytical data:

MS, m/e (rel. intensity, %): 231 (83) [M+H]⁺, 248 (100) [M+NH₄]⁺.

20 Example 30: N-(Cyanoacetyl)β-ethanolamide (A₁₁)

To β-ethanolamine (1.37 ml, 22.6 mmol), methyl cyanoacetate was added (2.0 ml, 22.6 mmol). The reaction was heated for 30 h at 100°C. Cooling gave a brown solid which was recrystallized from ethanol to give 2.10 g of the product (71%). The product gave the following analytical data: MS, m/e (rel. intensity, %):129 (30) [M+H]⁺, 146 (100) [M+NH₄]⁺.

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Exampl 31: (E,E)-2-(β-Ethanolamido)-3-(3,5-dimethoxy-4hydroxystyryl)acrylonitril (CR24)

To 3,5-dimethoxy-4-hydroxycinnamaldehyde (0.018 g, 0.086 mmol), N-(cyanoacetyl)β-ethanolamide (Example 30, 0.010 g, 0.086 mmol) was added. After refluxing for 2 h and purification on silica gel, CHCl₃-MeOH, 5:1, a yellow solid was obtained. (0.024 g, 87%). The product gave the following analytical data:

NMR (CD₃COCD₃, δ , ppm): 3.47, 3.67 (2 × m, 4H, NHCH₂ + CH₂OH), 3.90 (s, 6H, OCH₃ + OCH₃), 7.07 (br.s, 2H, H²⁺⁶), 7.16 (dd, 1H, J 11.7 and 15.2 Hz, 10 PhC=CH olefinic), 7.31 (br.s, 1H, NH), 7.38 (d, 1H, J 15.2 Hz, PhCH olefinic), 7.97 (d, 1H, J 11.7 Hz, CH=CCN olefinic).

MS, m/e (rel. intensity, %): 319 (70) $[M+H]^+$, 341 (100) $[M+Na]^+$.

Example 32: (E,E)-2-(Benzylamido)-3-(4-nitrostyryl)acrylonitrile (CR27) 15

The compound was prepared as described in Example 3 by adding 4nitrocinnamaldehyde (0.022 g, 0.12 mmol) to N-(cyanoacetyl)benzylamide (Example 4, 0.022 g, 0.12 mmol). After refluxing for 1 h, the product was purified by silica gel chromatography (CHCl3-MeOH, 5:1) to give a yellow solid (0.033 g, 81%). The product gave the following analytical data:

NMR (CD₃COCD₃, δ, ppm): 4.56 (br.s, 2H, NHCH₂), 7.24-7.38 (m, 6H, Ph' + NH), 7.47 (dd, 1H, J 11.1 and 15.2 Hz, PhC=CH olefinic), 7.62 (d, 1H, J 15.2 Hz, PhCH olefinic), 8.02, 8.32 (2 × br.d, 4H, J 8.8 and 8.3 Hz, Ph), 8.08 (d,

1H, J 11.1 Hz, CH=CCN olefinic). 25

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MS, m/e (rel. intensity, %): 334 (100) $[M+H]^+$, 351 (16) $[M+NH_4]^+$, 356 (28) $[M+Na]^+$.

Example 33: (E,E)-2-(3,4-Dihydroxybenzylamido)-3-(4-nitrostyryl)acrylonitrile(CR28)

The compound was prepared as described in Example 3 by adding 4-nitrocinnamaldehyde (0.009 g, 0.05 mmol) to N-(cyanoacetyl)3,4-dihydroxybenzylamide (Example 2, 0.010 g, 0.05 mmol). After refluxing for 2 h and recrystallization from ethanol a yellow solid was obtained (0.007g, 39%). The product gave the following analytical data:

NMR (CD₃COCD₃, δ , ppm): 2.81, 2.83 (2 × br.s, 2H, OH + OH), 4.39 (br.s, 2H, NHCH₂), 6.69 (br.d, 1H, J < 0.5 and 7.6 Hz, H⁶), 6.76 (d, 1H, J 7.6 Hz, H⁵), 6.86 (br.d, 1H, J < 0.5 Hz, H²), 7.47 (dd, 1H, J 11.7 and 15.2 Hz, PhC=CH olefinic), 7.61 (d, 1H, J 15.2 Hz, PhC olefinic), 7.91 (br.s, 1H, NH), 8.02, 8.31 (2 × br.d, 4H, J 8.2 and 8.8 Hz, Ph), 8.06 (d, 1H, J 11.7 Hz, CH=CCN olefinic).

MS, m/e (rel. intensity, %): 331 (21) [M-OH-OH]⁺, 348 (47) [M-OH]⁺, 366 (100) [M+H]⁺, 383 (97) [M+NH₄]⁺.

Example 34: (E,E)-2-(1-Amino-2,2-dicyanoethenyl)-3-(4-nitrostyryl)acrylonitrile (CR29)

The compound was prepared as described in Example 3 by adding 4-nitrocinnamaldehyde (0.051 g, 0.29 mmol) to 2-amino-1-propene-1,1,3-tricarbonitrile (0.038 g, 0.29 mmol). After refluxing for 4 h and recrystallization

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from ethanol a yellow solid was obtained (0.08 g, 51%). The product gave the following analytical data:

NMR (CD₃COCD₃, δ , ppm): 7.54 (dd, 1H, J 11.1 and 15.8 Hz, PhC=CH olefinic), 7.67 (d, 1H, J 15.8 Hz, PhCH olefinic), 7.99 (d, 1H, J 11.1 Hz, CH=CCN olefinic), 8.08, 8.32 (2 × br.d, 4H, J 8.8 and 8.8 Hz, Ph). MS, m/e (ref. intensity, %): 309 (100) [M+NH₄]⁺, 314 (67) [M+Na]⁺.

Example 35: Effect of CR4 Upon Normal Bone Marrow Differentiation in Culture

The CFU-GEMM assay was performed according to Fauser and Messner (1978, Blood, 52(6) 143-8) and Messner and Fausser (1980, Blut, 41(5) 327-33) with some variations. In brief, heparinized bone marrow cells were layered over Percoll (1.077 gm/ml) (Pharmacia Fine Chemical, Piscataway NJ) and centrifuged at 400g at 4°C for 10 minutes to remove neutrophils and RBCs. The fractionated BM cells at 2x10⁵ cells/ml were cultured in IMDM (OCI, Toronto) containing 0.9% (vol/vol) methylcellulose supplementd with 30% FCS (Cansera Rexdale, ON.) or normal human plasma, a cocktail of cytokines containing G-CSF (10 ng/ml, Amgen), IL-3 (40 U/ml, Immunex), MGF (50 ng/ml, Immunex), Erythropoietin (2u/ml, Epprex) or TPO (10 ng/ml, Amgen), 5x10⁻⁵M β-2-mercaptoethanol and the specified concentration of CR4. The culture mixture was plated in 1 ml volumes into 35 mm petri dishes and incubated at 37°C, 5% CO₂ in a humidified atmosphere. All cultures were evaluated at 14 days for the number of BFU-E colonies (defined as aggregates of more than 500 hemoglobinized cells or, 3 or more erythroid subcolonies), CFU-GM colonies (defined as granulocyte or monocyte-macrophage cells or both), CFU-Meg colonies (comprising 4 or more megakaryocytes) and CFU-GEMM colonies (a mixed population comprising of all elements).

The results shown in Figure 1 demonstrate that CR4 displayed negligible toxicity upon normal bone marrow at doses up to 5 μM. At 10 μM

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CR4 began to cause some inhibition of BFU-E colony formation, but at the same tim significantly stimulated CFU-GM colony numbers.

Example 36: Killing of Philadelphia positive Acute Lymphoblastic Leukemia by low-dose CR4 in culture.

Ph+ ALL cells were plated in 1 ml volumes, in the absence of exogenous growth factors, into 35 mm petri dishes (Nunc, Gibco) containing alpha MEM (Gibco) plus 10% FCS (Cansera Rexdale, ON.) in 0.9% (vol/vol) methylcellulose (Fluka, Switzerland) with the indicated concentrations of CR4. Cultures were set at 37°C, 5% CO₂ in a humidified atmosphere. Colonies consisting of more than 20 cells were counted at 12 days or earlier using an inverted microscope.

The results shown in Figure 2 demonstrate that CR4 effected a significant inhibition of Ph+ ALL cell proliferation and survival at low nanomolar doses (35-100). CR4 has no effect upon normal cells at equivalent concentrations.

Example 37: Killing of Philadelphia positive Z119 Acute Lymphoblastic Leukemia Cells by low-dose CR4 in culture.

Z119 cells were plated in 1 ml volumes at a density of 1x10⁴ cells/ml, in the absence of exogenous growth factors, into 35 mm petri dishes (Nunc, Gibco) containing IMDM (OCI, Toronto) plus 20% FCS (Cansera Rexdale, ON.) in 0.9% (vol/vol) methylcellulose (Fluka, Switzerland) with the indicated concentration of CR4. Cultures were set at 37°C, 5% CO₂ in a humidified atmosphere. Colonies consisting of more than 20 cells were counted at 7 days or earlier using an inverted microscope.

The results shown in Figure 3 demonstrate that CR4 effected a significant inhibition of Z119 ALL cell proliferation and survival at low nanomolar doses. CR4 has no effect upon normal cells at equivalent concentrations.

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Example 38: Killing of AML-3 Acute Myeloid Leukemia Cells by low-dose CR4 in culture

OCI-AML-3 cells were plated in 35mm petri dishes (Nunc, Gibco) in 1 ml volumes at a density of $3.3x10^3$ cells/ml, in the absence of exogenous growth factors, containing alpha MEM plus 20% FCS (Cansera, Rexdale Ont.), and 0.9% (vol/vol) methylcellulose (Fluka, Switzerland) and the indicated concentrations of CR4. Cell cultures were incubated in a humidified atmosphere at 37°C with 5% CO₂. Colonies containing more than 20 cells were scored, using an inverted microscope, at 5-6 days.

The results shown in Figure 4 demonstrate that CR4 effected a complete inhibition of AML-3 cell proliferation and survival at nanomolar concentrations (300-600nM). CR4 has no effect upon normal cell survival at equivalent concentrations.

15 Example 39: Killing of Ly-MN Lymphoma cells by low-dose CR4 in culture.

Ly-MN cells were plated in 1 ml volumes, in the absence of exogenous growth factors, into 35 mm petri dishes (Nunc, Gibco) containing IMDM (OCI, Toronto) plus 20% human cord blood plasma in 0.9% (vol/vol) methylcellulose (Fluka, Switzerland) and the indicated concentrations of CR4. Cultures were set at 37°C, 5% CO₂ in a humidified atmosphere. Colonies consisting of more than 20 cells were counted at 5 days or earlier using an inverted microscope.

The results shown in Figure 5 demonstrate that CR4 significantly inhibited cell proliferation and survival at nanomolar doses, and effected a inhibition by 2.5 μ M. CR4 has no effect upon normal cells at equivalent concentrations.

Example 40: Killing of Primary Juvenile Myelo-Monocytic Leukemia Cells by CR4 in Culture.

Heparinized bone marrow cells from a JMML patient were layered over Percoll (1.077 gm/ml) (Pharmacia Fine Chemical, Piscataway NJ) and centrifuged at 400g at 4°C for 10 minutes to remove neutrophils and RBCs.

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Cells were further fractionated and purified on Miltenyi MS columns (Miltenyi Biotec GmbH, Germany) to acquire an early progenitor population of CD34⁺ cells. The fractionated BM CD34⁺ cells at a density of 1 x 10⁴ cells/ml were cultured in IMDM (OCI, Toronto) containing 0.9% (vol/vol) methylcellulose supplemented with 30% FCS (Cansera Rexdale, ON.) with the indicated concentration of CR4. The culture mixture was plated in 1 ml volumes into 35 mm petri dishes and incubated at 37°C, 5% CO₂ in a humidified atmosphere. Colonies consisting of more than 20 cells were counted at 12 days or earlier using an inverted microscope.

The results shown in Figure 6 demonstrate that CR4 displayed moderate killing ability with primary JMML cells, with 80-90 percent inhibition achieved by $5 \mu M$ concentrations.

Example 41: Killing of OCI-LY2 Lymphoma Cells by low-dose CR4 in culture.

OCI-LY2 cells were plated in 1 ml volumes, in the absence of exogenous growth factors, into 35 mm petri dishes (Nunc, Gibco) containing IMDM (OCI, Toronto) plus 20% human cord blood plasma in 0.9% (vol/vol) methylcellulose (Fluka, Switzerland) and the indicated doses of CR4. Cultures were set at 37°C, 5% CO₂ in a humidified atmosphere. Colonies consisting of more than 20 cells were counted at 5 days or earlier using an inverted microscope.

The results shown in Figure 7 demonstrate that CR4 significantly inhibited cell proliferation and survival at high nanomolar to low micromolar doses (90% at 2.5 μ M). CR4 has no effect upon normal cells at equivalent concentrations.

Example 42: Killing of Philadelphia positive ALL Cells by CR17 and CR21 in culture.

ALL cells were plated in 1 ml volumes, in the absence of exogenous growth factors, into 35 mm petri dishes (Nunc, Gibco) containing alpha MEM (Gibco) plus 20% FCS (Cansera Rexdale, ON.) in 0.9% (vol/vol)

methylcellulose (Fluka, Switzerland) and the indicated concentrations of compound. Cultures were set at 37°C, 5% CO₂ in a humidified atmosphere. Colonies consisting of more than 20 cells were counted at 12 days or earlier using an inverted microscope.

The results shown in Figure 8 demonstrate that CR17 displayed significant inhibition of cell growth at 1-2.5 μ M concentrations. CR21 inhibited cell growth at 5 μ M.

Example 43: Killing of Philadelphia positive ALL Cells by CR17 and CR21 in culture.

ALL cells were plated in 1 ml volumes, in the absence of exogenous growth factors, into 35 mm petri dishes (Nunc, Gibco) containing alpha MEM (Gibco) plus 20% FCS (Cansera Rexdale, ON.) in 0.9% (vol/vol) methylcellulose (Fluka, Switzerland) and the indicated concentrations of compound. Cultures were set at 37°C, 5% CO₂ in a humidified atmosphere. Colonies consisting of more than 20 cells were counted at 12 days or earlier using an inverted microscope.

The results shown in Figure 9 demonstrate that CR17 and CR21 both displayed significant inhibition of cell growth at 1-2.5 μ M concentrations.

Example 44: Killing of Philadelphia positive ALL Cells by CR24 in culture.

ALL cells were plated in 1 ml volumes, in the absence of exogenous growth factors, into 35 mm petri dishes (Nunc, Gibco) containing alpha MEM (Gibco) plus 20% FCS (Cansera Rexdale, ON.) in 0.9% (vol/vol) methylcellulose (Fluka, Switzerland) and the indicated concentrations of CR24. Cultures were set at 37°C, 5% CO₂ in a humidified atmosphere. Colonies consisting of more than 20 cells were counted at 9 days or earlier using an inverted microscope.

The results shown in Figure 10 demonstrate that CR24 was effective against Ph+ ALL cells at concentrations as low as 0.5 μ M, demonstrating a virtually complete inhibition of cell growth between 2.5 and 5 μ M.

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Exampl 45: Killing of Philadelphia positive ALL Cells by CR19 in culture.

ALL cells were plated in 1 ml volumes, in the absence of exogenous growth factors, into 35 mm petri dishes (Nunc, Gibco) containing alpha MEM (Gibco) plus 20% FCS (Cansera Rexdale, ON.) in 0.9% (vol/vol) methylcellulose (Fluka, Switzerland) and the indicated concentrations of CR19. Cultures were set at 37°C, 5% CO₂ in a humidified atmosphere. Colonies consisting of more than 20 cells were counted at 9 days or earlier using an inverted microscope.

The results shown in Figure 11 demonstrate that CR19 was highly effective against Ph+ ALL cells at nanomolar concentrations between 250 and 500nM.

15 Example 46: Effect of CR19 on Normal Bone Marrow Differentiation in Culture.

The CFU-GEMM assay was performed according to Fauser and Messner (1978, Blood, 52(6) 1243-8) and Messner and Fausser (1980, Blut, 41(5) 327-33) with some variations. In brief, heparinized bone marrow cells were layered over Percoll (1.077 gm/ml) (Pharmacia Fine Chemical, Piscataway NJ) and centrifuged at 400g at 4°C for 10 minutes to remove neutrophils and RBCs. The fractionated BM cells at 2x10⁵ cells/ml were cultured in IMDM (OCI, Toronto) containing 0.9% (vol/vol) methylcellulose supplementd with 30% FCS (Cansera Rexdale, ON.) or normal human plasma, a cocktail of cytokines containing G-CSF (10 ng/ml, Amgen), IL-3 (40 U/ml, Immunex), MGF (50 ng/ml, Immunex), Erythropoietin (2u/ml, Epprex) or TPO (10 ng/ml, Amgen), 5x10⁻⁵M β-2-mercaptoethanol and the specified concentrations of CR19. The culture mixture was plated in 1 ml volumes into 35 mm petri dishes and incubated at 37°C, 5% CO₂ in a humidified atmosphere. All cultures were evaluated at 14 days for the number of BFU-E colonies (defined as aggregates of more than 500 hemaglobinized cells or, 3

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or more erythroid subcolonies) and CFU-C colonies (d fined as granulocyte or monocyte-macrophage cells or both).

The results shown in Figure 12 demonstrate that CR19 displayed significant inhibition of the development of BFU-E colonies at 2.5 μ M, although at this concentration it also boosted CFU-C colony formation. At 5 μ M the stimulatory effect disappeared and BFU-E colonies were virtually absent.

Example 47: Effect of CR24, CR17 and CR21 on Normal Bone Marrow Differentiation.

The CFU-GEMM assay was performed according to Fauser and Messner (1978, Blood, 52(6) 1243-8) and Messner and Fausser (1980, Blut, 41(5) 327-33) with some variations (British Journal of Haematology, 1992, 80, p40-48). In brief, heparinized bone marrow cells were layered over Percoll (1.077 gm/ml) (Pharmacia Fine Chemical, Piscataway NJ) and centrifuged at 400g at 4°C for 10 minutes to remove neutrophils and RBCs. The fractionated BM cells at 2x105 cells/ml were cultured in IMDM (OCI, Toronto) containing 0.9% (vol/vol) methylcellulose supplementd with 30% FCS (Cansera Rexdale, ON.) or normal human plasma, a cocktail of cytokines containing G-CSF (10 ng/ml, Amgen), IL-3 (40 U/ml, Immunex), MGF (50 ng/ml, Immunex), Erythropoietin (2u/ml, Epprex) or TPO (10 ng/ml, Amgen), 5x10⁻⁵M β-2mercaptoethanol and the specified concentration of test compound. The culture mixture was plated in 1 ml volumes into 35 mm petri dishes and incubated at 37°C, 5% CO2. In a humidified atmosphere. All cultures were evaluated at 14 days for the number of BFU-E colonies (defined as aggregates of more than 500 hemoglobinized cells or, 3 or more erythroid subcolonies) and CFU-C colonies (defined as granulocyte or monocytemacrophage cells or both).

The results shown in Figure 13 demonstrate that CR24 displayed minimal inhibition of bone marrow colony formation at either 10 or 20 μM

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concentrations, whereas both CR17 and CR21 caused inhibition of BFU-E colony formation at the higher 20 μ M dose.

Example 48: In vitro Purging of Normal Bone Marrow with CR4.

Heparinized bone marrow cells were layered over Percoll (1.077 gm/ml) (Pharmacia Fine Chemical, Piscataway NJ) and centrifuged at 400g at 4°C for 10 minutes to remove neutrophils and RBCs.

For the purging process, the cells were resuspended at 1x10⁶/ml in complete medium with or without 50 µM CR4. The cells were incubated with the CR4 for two and a half hours at 37°C, 5% CO2. At the end of this period the cells were thoroughly washed in medium to remove CR4 and then cultured at 2x10⁵ cells/ml in IMDM (OCI, Toronto) containing 0.9% (vol/vol) methylcellulose supplemented with 30% FCS (Cansera Rexdale, ON.) or normal human plasma, a cocktail of cytokines containing G-CSF (10 ng/ml, Amgen), IL-3 (40 U/ml, Immunex), MGF (50 ng/ml, Immunex), Erythropoietin (2u/ml, Epprex) or TPO (10 ng/ml, Amgen) and 5x10⁻⁵M b-2-mercaptoethanol. The culture mixture was plated in 1 ml volumes into 35 mm petri dishes and incubated at 37°C, 5% CO2 in a humidified atmosphere. All cultures were evaluated at 14 days for the number of BFU-E colonies (defined as aggregates of more than 500 hemaglobinized cells or, 3 or more erythroid subcolonies), CFU-C colonies (defined as granulocyte or monocytemacrophage cells or both) and CFU-GEMM colonies (a mixed population comprising of all elements).

The results shown in Figure 14 demonstrate that two and a half hours exposure to 50 μ M CR4 did not result in significant inhibition of colony formation. While a slight drop in BFU-E colonies occurred, CFU-C colony numbers actually increased significantly.

Example 49: In Vitro Purging of Z119 Acute Lymphoblastic Leukemia with CR4.

For the purging assay, the cells were resuspended in complete medium with or without CR4 as indicated and incubated at 37°C, 5%CO₂ for

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0-5 hours. C IIs were then washed thoroughly with medium to remove the CR4, resuspended and plated in 1 ml volumes, in the absence of exogenous growth factors, into 35 mm p. tri dishes (Nunc, Gibco) containing alpha MEM (Gibco) plus 20% FCS (Cansera Rexdale, ON.) in 0.9% (vol/vol) methylcellulose (Fluka, Switzerland). Cultures were set at 37°C, 5% CO₂ in a humidified atmosphere. Colonies consisting of more than 20 cells were counted at 9 days or earlier using an inverted microscope.

The results shown in Figure 15 demonstrate that CR4 demonstrated rapid killing of Z119 cells at the concentrations examined. 50 μ M CR4 displayed a complete inhibition after only 2.5 hours exposure, while 85% killing could be achieved with 25 μ M CR4 over the same time period. A longer five hour exposure of the cells to 25 μ M CR4 resulted in a complete ablation of subsequent cell growth.

15 Example 50: In vitro Purging of OCI-Ly2 Lymphoma cells with CR4.

For the purging assay, the cells were resuspended in complete medium with or without CR4 as indicated and incubated at 37°C, 5%CO₂ for 0-5 hours. Cells were then washed thoroughly with medium to remove CR4, resuspended and plated in 1 ml volumes at 5x10³ cellls/ml, in the absence of exogenous growth factors, into 35 mm petri dishes (Nunc, Gibco) containing IMDM (OCI, Toronto) plus 20% human cord blood plasma in 0.9% (vol/vol) methylcellulose (Fluka, Switzerland). Cultures were set at 37°C, 5% CO₂ in a humidified atmosphere. Colonies consisting of more than 20 cells were counted at 5 days or earlier using an inverted microscope.

The results shown in Figure 16 demonstrate that CR4 demonstrated significant killing (90%) of OCI-Ly2 cells at 25-50 μ M after only 5 hours exposure. The lower 12.5 μ M dose tested also achieved significant killing in the same time period.

30 Example 51: *In vitro* Purging of OCI-AML-3 Acute Meyloid Leukemia Cells with CR4.

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For the purging assay, the cells were resuspended in complete medium with or without CR4 as indicated and incubated at 37°C, 5%CO₂ for 0-5 hours. Cells were then washed thoroughly with medium to remove CR4, resuspended and plated in 1 ml volumes at 5x10³ cells/ml, in the absence of exogenous growth factors, into 35 mm petri dishes (Nunc, Gibco) containing alpha MEM (Gibco) plus 10% FCS (Cansera Rexdale, ON.) in 0.9% (vol/vol) methylcellulose (Fluka, Switzerland). Cultures were set at 37°C, 5% CO₂ in a humidified atmosphere. Colonies consisting of more than 20 cells were counted at 5 days or earlier using an inverted microscope.

The results shown in Figure 17 demonstrate that CR4 demonstrated significant killing of OCI-AML-3 cells at 50 μ M after only 2.5-5 hours exposure. The lower 25 μ M dose tested also achieved significant killing in the same time period.

15 Example 52: In vitro Purging of Ramos B Cell Burkitt's Lymphoma Cells with CR4.

For the purging assay, the cells were resuspended in complete medium with or without CR4 as indicated and incubated at 37°C, 5%CO₂ for 0-5 hours. Cells were then washed thoroughly with medium to remove the CR4, resuspended and plated in 1 ml volumes, in the absence of exogenous growth factors, into 35 mm petri dishes (Nunc, Gibco) containing RPMI 1640 (Gibco) plus 10% FCS (Cansera Rexdale, ON.) in 0.9% (vol/vol) methylcellulose (Fluka, Switzerland). Cultures were set at 37°C, 5% CO₂ in a humidified atmosphere. Colonies consisting of more than 20 cells were counted at 12 days or earlier using an inverted microscope.

The results shown in Figure 18 demonstrate that CR4 demonstrated significant killing (70%) of Ramos cells at 50 µM after 5 hours exposure.

Example 53: Killing of HuNS1 multiple myeloma by CR4.

HuNS1 cells were plated in 35mm petri dishes (Nunc, Gibco) in 1ml volumes at a density of 1x10⁴ cells/ml, in the absence of exogenous growth factors, containing alpha MEM plus 20% FCS (Cansera, Rexdale Ont.), and

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0.9% (vol/vol) methylcellulose (Fluka, Switzerland) and the indicated concentrations of CR4. Cell cultures were incubated in a humidified atmosphere at 37°C with 5% CO₂. Colonies containing more than 20 cells were scored, using an inverted microscope, at 5-6 days.

The results shown in Figure 19 demonstrate that CR4 significantly inhibited cell proliferation and survival at high nanomolar to low micromolar doses (>90% at 2.5μM). CR4 has no effect upon normal cells at equivalent concentrations.

Example 54: In Vivo Treatment of Philadelphia positive Acute Lymphoblastic Leukemia in NOD-SCID mice.

NOD-SCID mice were irradiated (350 rads) and injected with 5x10⁶ Philadelphia positive Z119 Acute lymphoblastic leukemia cells. After 24 hours Alzet micro-osmotic pumps (Alza Corp. Paolo Alto, CA) were implanted subcuntaneously, containing either 20 mM solution of CR4 in 50% DMSO/medium or 50% DMSO/medium alone. Alzet 2001 pumps were utilized, holding a total volume of 200µl and releasing 1µl per hour over 7-10 days. Pumps were replaced after 7 days. Each mouse received a daily dose of 0.154 mg of CR4.

After 14 (Fig.20A) and 21 (Fig.20B) days mice were sacrificed and bone marrow extracted from the fore and hind limbs. Single cell suspensions were prepared, red blood cells lysed and the samples stained with PE-labelled isotype, anti-human CD19 and anti-human HLA-DR antibodies to detect the presence of Z119 cells. These antibodies do not cross react with murine cells.

At d14, bone marrow cell cultures were also performed to assess the presence of Z119 cells. $5x10^4$ BM cells were cultured in IMDM (OCI, Toronto) containing 0.9% (vol/vol) methylcellulose supplemented with 30% serum consisting of a 1:1 mixture of FCS (Cansera Rexdale, ON.) and normal human plasma. No cytokines are added. Under these conditions there is no growth of murine cells. The culture mixture was plated in 1 ml volumes into 35 mm petri dishes and incubated at 37°C, 5% CO₂ in a humidified atmosphere. All cultures were evaluated at 9 days for the number of ALL colonies.

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The results shown in Figures 20A and 20B demonstrate that at both day 14 and 21 sacrifices, a significant reduction in ALL infiltration of the bone marrow was observed in all the mice treated with CR4 relative to the DMSO treated control mice.

In control animals, massive infiltration of the spleen, liver and kidney was observed, as well as the presence of ALL cells in the peripheral blood. In addition to a 90% reduction in the infiltration of ALL cells into the bone marrow, treatment with CR4 reduced ALL infiltration of the organs and blood to below detectable levels.

Thus, CR4 was highly effective against a variety of cancer cells, including acute lymphoblastic leukemia, Philadelphia positive ALL, acute myeloid leukemia, myeloma and B-lineage lymphoma, at concentrations ranging from 50nM to 5μM. At the same time, minimal toxicity was seen when normal cells were incubated in the presence of CR4 until concentrations of 10-20μM or greater were achieved. CR4 was particularly active against bcrabl transformed Philadelphia positive cells, achieving >90% wipeout at concentrations as low as 40nM. CR4 was also highly effective in high dose (25-50μM) *in vitro* purging assays against Philadelphia positive ALL, AML and lymphoma, causing >90% inhibition of growth with a 2.5 to 5 hour exposure time. Over identical doses and times normal bone marrow growth and differentiation were unaffected. CR4 showed a combination of high level toxicity to cancer cells with minimal non-specific cytotoxic damage.

CR4 was also highly effective in a whole animal model (Example 54). The compound demonstrated good retention characteristics, still being detectable in the blood 30 minutes after I.V. injection. In a murine model of human Ph+ ALL, CR4 caused a greater than 90 percent reduction in ALL infiltration of bone marrow within a two week period, reducing the presence of infiltrating ALL cells in liver, kidney, spleen and peripheral blood below detection level. In contrast, control mice treated with vehicle alone demonstrated massive infiltration of all these organs. No evidence of non-specific toxicity was observed.

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Example 55: In vitro Purging of Normal Bone Marrow with CR11.

Heparinized bone marrow cells were layered over Percoll (1.077 gm/ml) (Pharmacia Fine Chemical, Piscataway NJ) and centrifuged at 400g at 4°C for 10 minutes to remove neutrophils and RBCs.

For the purging process, the cells were resuspended at 1x10⁸/ml in complete medium with or without 50µM CR11. The cells were incubated with the tryrphostin for seven hours at 37°C, 5% CO₂. At the end of this period the cells were thoroughly washed in medium to remove CR11 and then cultured at 2x10⁵ cells/ml in IMDM (OCI, Toronto) containing 0.9% (vol/vol) methylcellulose supplementd with 30% FCS (Cansera Rexdale, ON.) or normal human plasma, a cocktail of cytokines containing G-CSF (10 ng/ml, Amgen), IL-3 (40 U/ml, Immunex), MGF (50 ng/ml, Immunex), Erythropoietin (2u/ml, Epprex) or TPO (10 ng/ml, Amgen) and 5x10⁻⁵M β-2-mercaptoethanol. The culture mixture was plated in 1 ml volumes into 35 mm petri dishes and incubated at 37°C, 5% CO2 in a humidified atmosphere. All cultures were evaluated at 14 days for the number of BFU-E colonies (defined as aggregates of more than 500 hemoglobinized cells or, 3 or more erythroid subcolonies), CFU-C colonies (defined as granulocyte or monocytemacrophage cells or both) and CFU-GEMM colonies (a mixed population comprising of all elements).

The results shown in Figure 21 demonstrate that seven hours exposure to 50µM CR11 did not result in any significant inhibition of colony formation. BFU-E, CFU-GEMM and CFU-C colonies were all normal.

25 Example 56: In Vitro Purging of Philadelphia positive Acute Lymphoblastic Leukemia with CR11.

For the purging assay, the cells were resuspended in complete medium with or without CR11 as indicated and incubated at 37°C, 5%CO₂ for 0-7 hours. Cells were then washed thoroughly with medium to remove the CR11, resuspended and plated in 1 ml volumes, in the absence of exogenous growth factors, into 35 mm petri dishes (Nunc, Gibco) containing alpha MEM (Gibco) plus 20% FCS (Cansera Rexdale, ON.) in 0.9% (vol/vol)

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methylcellulose (Fluka, Switzerland). Cultures were set at 37°C, 5% CO₂ in a humidified atmosphere. Colonies consisting of more than 20 cells were counted at 9 days or earlier using an inverted microscope.

The results shown in Figure 22 demonstrate that CR11 demonstrated complete killing of Ph+ ALL cells at 50µM after 7 hours exposure.

Example 57: Philadelphia (Ph+) ALL lines Z119 and Z181 (5x10⁸ cells/point) were lysed and immunoprecipitated with Bcr-Abl antibody.

The precipitates were washed twice with lysis buffer and once with kinase assay buffer, and resuspended in same buffer containing varying concentrations of CR4. The precipitates were incubated with the drug for 10 min at room temperature, followed by addition of 10µCi ³³PγATP. The reaction was stopped after 20 min by the addition of SDS-PAGE reducing sample buffer and separated on an 8-16% SDS-PAGE gel. The products were transferred onto nitrocellulose membrane and visualized by autoradiography.

The results shown in Figure 23 demonstrate that Bcr-Abl kinase activity is effectively blocked at concentrations of 1 to 10 μ M of the CR4 compound in both Z199 and Z181 ALL cell lines.

20 Example 58: Philadelphia (Ph+) ALL line Z119 (5x10⁶ cells/point) was preincubated for 5 hours with different concentrations of CR4 and immunoprecipitated with Jak2 antibody.

The cells were lysed in lysis buffer and immunoprecipitated with Jak2 antibody. The precipitates were washed twice with lysis buffer and once with kinase assay buffer, followed by addition of 10μCi ³³PγATP. The reaction was stopped after 20 min by the addition of SDS-PAGE reducing sample buffer and separated on an 8-16% SDS-PAGE gel. The products were transferred onto nitrocellulose membrane and visualized by autoradiography.

The results shown in Figure 24 demonstrate that Jak2 kinase activity was dramatically inhibited at a concentration of $6\mu M$ and further blocked at higher concentrations.

While the present invention has been described with reference to what are presently considered to be the preferred examples, it is to be understood that the invention is not limited to the disclosed examples. To the contrary, the invention is intended to cover various modifications and equivalent arrangements included within the spirit and scope of the appended claims.

All publications, patents and patent applications are herein incorporated by reference in their entirety to the same extent as if each individual publication, patent or patent application was specifically and individually indicated to be incorporated by reference in its entirety.